

2021

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10.1128/MRA.00599-21

O'Grady, K., Riley, T. V., & Knight, D. R. (2021). Complete genome assemblies of three highly prevalent, toxigenic clostridioides difficile strains causing health care-associated infections in Australia. *Microbiology Resource Announcements*, 10(31), article e00599-21. <https://doi.org/10.1128/MRA.00599-21>

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# Complete Genome Assemblies of Three Highly Prevalent, Toxigenic *Clostridioides difficile* Strains Causing Health Care-Associated Infections in Australia

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**ABSTRACT** *Clostridioides difficile* infection (CDI) is the leading cause of life-threatening health care-related gastrointestinal illness worldwide. Phylogenetically appropriate closed reference genomes are essential for studies of *C. difficile* transmission and evolution. Here, we provide high-quality complete hybrid genome assemblies for the three most prevalent *C. difficile* strains causing CDI in Australia.

*Clostridioides difficile* causes life-threatening diarrhea and health care-related gastrointestinal infections globally (1). Core genome single nucleotide polymorphism (cgSNP) analysis is highly discriminatory for bacterial transmission and outbreak detection studies and the gold standard for reconstructing large phylogenies of closely related microbes (2). A critical step in cgSNP analysis involves mapping raw sequence data to a closely related reference genome, allowing for variant sites to be identified, filtered, and compared between strains (3). Using phylogenetically related “closed” reference genomes provides optimal mapping and variant calling. Australia has a diverse *C. difficile* population distinct from that of the rest of the world (1, 4), yet there are no phylogenetically appropriate reference genomes. Here, combining short- and long-read sequence technologies, we provide high-quality complete genome sequences for three of the most prevalent *C. difficile* strains causing *C. difficile* infection (CDI) in Australia, PCR ribotype 014 (RT014) (29.5% prevalence), RT002 (11.8%), and RT056 (5.4%) (5, 6). Representative *C. difficile* strains of each ribotype (S-0352, S-0253, and S-0942, respectively) were selected from >1,500 isolates recovered from patients with symptomatic CDI, part of the ongoing nationwide longitudinal surveillance of CDI in Australia, the *C. difficile* Antimicrobial Resistance Surveillance (CDARS) study (5, 6).

*C. difficile* strains from CDARS were cultured on blood agar in an anaerobic chamber (80% N<sub>2</sub>, 10% CO<sub>2</sub>, 10% H<sub>2</sub>) for 48 h (5). Total genomic DNA was extracted using a QuickGene DNA tissue kit (Kurabo Industries, Osaka, Japan) and used as input for both short-read (Illumina) and long-read (Oxford Nanopore Technologies [ONT]) sequencing. Illumina whole-genome sequencing (WGS) was performed using standard Nextera Flex paired-end read (2 × 150-bp) libraries on an Illumina NovaSeq 6000 instrument (Illumina, San Diego, CA, USA) to an average read depth of 130×. Default parameters were used for all software unless specified. The raw reads were filtered for quality (Q30+) and adaptor sequences using Trim Galore v0.6.5 (<https://github.com/FelixKrueger/TrimGalore>). ONT sequencing was performed on a MinION Mk1C device (ONT, Oxford, UK) using an R9 generation flow cell following a DNA by ligation protocol (SQK-LSK109). Filtlong v0.2.0 (<https://github.com/rwick/Filtlong>) was used to filter the low-quality reads (keeping the top 90% of reads and removing reads of <1,000 bp), resulting in 2.28 (S-0352), 2.59 (S-0253), and 6.66 (S-0942) Gb of sequence data, respectively. The hybrid assembly of ONT and Illumina reads was performed using Unicycler v0.4.8 (7) with multiple rounds of polishing (Pilon v1.2.4,

**Citation** O'Grady K, Riley TV, Knight DR. 2021. Complete genome assemblies of three highly prevalent, toxigenic *Clostridioides difficile* strains causing health care-associated infections in Australia. *Microbiol Resour Announc* 10:e00599-21. <https://doi.org/10.1128/MRA.00599-21>.

**Editor** David Rasko, University of Maryland School of Medicine

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**Received** 15 June 2021

**Accepted** 9 July 2021

**Published** 5 August 2021

**TABLE 1** Key features of *C. difficile* genomes

| Feature                                | Data for strain:                               |   |  |
|--|--|---|--|
|  | S-0352   | S-0253  | S-0942   |
| Strain epidemiology <sup>a</sup>       | RT014, ST2, clade 1                            | RT002, ST8, clade 1                                 | RT056, ST34, clade 1                           |
| Toxin profile <sup>b</sup>             | A <sup>+</sup> B <sup>+</sup> CDT <sup>-</sup> | A <sup>+</sup> B <sup>+</sup> CDT <sup>-</sup>      | A <sup>+</sup> B <sup>+</sup> CDT <sup>-</sup> |
| Origin <sup>c</sup>                    | Human, CDI, VIC 2014                           | Human, CDI, SA 2014                                 | Human, CDI, SA 2016                            |
| GenBank accession no.                  | <a href="#">CP076377</a>                       | <a href="#">CP076401</a> , <a href="#">CP076402</a> | <a href="#">CP076376</a>                       |
| ENA accession no.                      | <a href="#">ERS5447138</a>                     | <a href="#">ERS5447236</a>                          | <a href="#">ERS5447376</a>                     |
| Genome size (bp)                       | 4,251,987                                      | 4,089,134 (4,095,894 <sup>d</sup> )                 | 4,129,159                                      |
| %GC                                    | 28.96  | 28.52   | 28.71  |
| No. of CDS <sup>e</sup>                | 3,790  | 3,591   | 3,648  |
| No. of contigs                         | 1  | 2   | 1  |
| No. of tRNAs                           | 90   | 90  | 90   |
| No. of rRNAs                           | 35   | 35  | 35   |
| No. of CRISPRs <sup>f</sup>            | 10   | 4   | 9  |
| Read metrics                           |  |   |  |
| Total no. of ONT reads                 | 545,760  | 760,250   | 1,400,000                                      |
| Average ONT read length (bp, filtered) | 5,518  | 4,663   | 9,038  |
| Total no. of Illumina reads (trimmed)  | 2,015,674                                      | 1,930,928   | 1,868,362                                      |

<sup>a</sup> RT, PCR ribotype; ST, multilocus sequence type.

<sup>b</sup> Presence/absence of full-length *tcdA*, *tcdB* (pathogenicity locus, PaLoc), and binary toxin *cdtA/B* (binary toxin locus, CdtLoc). CDT, *C. difficile* binary toxin.

<sup>c</sup> VIC, Victoria; SA, South Australia.

<sup>d</sup> Combined chromosome and plasmid length.

<sup>e</sup> CDS, coding sequences.

<sup>f</sup> CRISPRs, clustered regularly interspaced short palindromic repeats.

Racon v1.4.3) to improve the contiguity. Complete circular genomes were confirmed using Bandage v0.8.1 (8) and rotated to *dnaA* using Unicycler. The genomes were evaluated using QUAST v2.344 (<http://quast.sourceforge.net/quast>) and annotated using the NCBI Prokaryotic Genome Annotation Pipeline v5.2 (9). The multilocus sequence type (ST) was determined using PubMLST (10).

The summary genome features and metrics are shown in Table 1. A single 6,760-bp plasmid was identified in S-0253 (RT002). This data set increases the diversity of complete reference genomes available to the *C. difficile* research community, aiding future studies of *C. difficile* transmission and evolution.

**Data availability.** The genome data are available at GenBank under BioProject accession number [PRJNA734443](#) (complete genome assemblies) and at the ENA under BioProject accession number [PRJEB41588](#) (Illumina sequence data); see Table 1 for details.

## ACKNOWLEDGMENTS

K.O. was funded by an Australian Government Research Training Program Scholarship. D.R.K. was funded by a fellowship from the National Health and Medical Research Council (APP1138257).

## REFERENCES

- Knight DR, Elliott B, Chang BJ, Perkins TT, Riley TV. 2015. Diversity and evolution in the genome of *Clostridium difficile*. *Clin Microbiol Rev* 28:721–741. <https://doi.org/10.1128/CMR.00127-14>.
- Eyre DW, Walker AS. 2013. *Clostridium difficile* surveillance: harnessing new technologies to control transmission. *Expert Rev Anti Infect Ther* 11:1193–1205. <https://doi.org/10.1586/14787210.2013.845987>.
- Olson ND, Lund SP, Colman RE, Foster JT, Sahl JW, Schupp JM, Keim P, Morrow JB, Salit ML, Zook JM. 2015. Best practices for evaluating single nucleotide variant calling methods for microbial genomics. *Front Genet* 6:235. <https://doi.org/10.3389/fgene.2015.00235>.
- Dingle KE, Elliott B, Robinson E, Griffiths D, Eyre DW, Stoesser N, Vaughan A, Golubchik T, Fawley WN, Wilcox MH, Peto TE, Walker AS, Riley TV, Crook DW, Didelot X. 2014. Evolutionary history of the *Clostridium difficile* pathogenicity locus. *Genome Biol Evol* 6:36–52. <https://doi.org/10.1093/gbe/evt204>.
- Knight DR, Giglio S, Huntington PG, Korman TM, Kotsanas D, Moore CV, Paterson DL, Prendergast L, Huber CA, Robson J, Waring L, Wehrhahn MC, Weldhagen GF, Wilson RM, Riley TV. 2015. Surveillance for antimicrobial resistance in Australian isolates of *Clostridium difficile*, 2013–14. *J Antimicrob Chemother* 70:2992–2999. <https://doi.org/10.1093/jac/dkv220>.
- Hong S, Putsathit P, George N, Hemphill C, Huntington PG, Korman TM, Kotsanas D, Lahra M, McDougall R, Moore CV, Nimmo GR, Prendergast L, Robson J, Waring L, Wehrhahn MC, Weldhagen GF, Wilson RM, Riley TV, Knight DR. 2020. Laboratory-based surveillance of *Clostridium difficile* infection in Australian healthcare and community settings, 2013 to 2018. *J Clin Microbiol* 58:e01552-20. <https://doi.org/10.1128/JCM.01552-20>.

7. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
8. Wick RR, Schultz MB, Zobel J, Holt KE. 2015. Bandage: interactive visualization of de novo genome assemblies. *Bioinformatics* 31:3350–3352. <https://doi.org/10.1093/bioinformatics/btv383>.
9. Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
10. Griffiths D, Fawley W, Kachrimanidou M, Bowden R, Crook DW, Fung R, Golubchik T, Harding RM, Jeffery KJM, Jolley KA, Kirton R, Peto TE, Rees G, Stoesser N, Vaughan A, Walker AS, Young BC, Wilcox M, Dingle KE. 2010. Multilocus sequence typing of *Clostridium difficile*. *J Clin Microbiol* 48:770–778. <https://doi.org/10.1128/JCM.01796-09>.