Edith Cowan University [Research Online](https://ro.ecu.edu.au/)

[Research outputs 2022 to 2026](https://ro.ecu.edu.au/ecuworks2022-2026)

1-1-2023

Lepidium meyenii walp (maca)-derived extracellular vesicles ameliorate depression by promoting 5-HT synthesis via the modulation of gut–brain axis

Rui Hong

Lan Luo

Liang Wang Edith Cowan University

Zhao-Li Hu

Qi-Rong Yin

See next page for additional authors

Follow this and additional works at: [https://ro.ecu.edu.au/ecuworks2022-2026](https://ro.ecu.edu.au/ecuworks2022-2026?utm_source=ro.ecu.edu.au%2Fecuworks2022-2026%2F2782&utm_medium=PDF&utm_campaign=PDFCoverPages)

Part of the Medicine and Health Sciences Commons

[10.1002/imt2.116](http://dx.doi.org/10.1002/imt2.116)

Hong, R., Luo, L., Wang, L., Hu, Z. L., Yin, Q. R., Li, M., . . . Qi, S. H. (2023). Lepidium meyenii walp (maca)-derived extracellular vesicles ameliorate depression by promoting 5-HT synthesis via the modulation of gut–brain axis. iMeta, 2(3), article e116. . <https://doi.org/10.1002/imt2.116> This Journal Article is posted at Research Online. https://ro.ecu.edu.au/ecuworks2022-2026/2782

Authors

Rui Hong, Lan Luo, Liang Wang, Zhao-Li Hu, Qi-Rong Yin, Ming Li, Bin Gu, Bin Wang, Tao Zhuang, Xin-Yue Zhang, Yuan Zhou, Wan Wang, Lin-Yan Huang, Bing Gu, and Su-Hua Qi

This journal article is available at Research Online: <https://ro.ecu.edu.au/ecuworks2022-2026/2782>

RESEARCH ARTICLE

iMeta WILEY

Lepidium meyenii Walp (Maca)‐derived extracellular vesicles ameliorate depression by promoting 5‐HT synthesis via the modulation of gut–brain axis

Rui Hong^{1,2} | Lan Luo¹ | Liang Wang^{3,4,5} | Zhao-Li Hu⁶ | Qi-Rong Yin^{2,7} | Ming Li^1 | Bin Gu⁴ | Bin Wang⁷ | Tao Zhuang² | Xin-Yue Zhang² | Yuan Zhou¹ | Wan Wang¹ | Lin-Yan Huang¹ | Bing Gu^{1,[3](http://orcid.org/0000-0001-7764-3213)} | Su-Hua Qi^{1,[2](http://orcid.org/0000-0002-1479-701X)} | ©

1 School of Medical Technology, Xuzhou Medical University, Xuzhou, China

2 School of Pharmacy, Xuzhou Medical University, Xuzhou, China

3 Laboratory Medicine, Guangdong Provincial People's Hospital, Guangzhou, China

4 School of Medical Informatics and Engineering, Xuzhou Medical University, Xuzhou, China

5 Centre for Precision Health, School of Medical and Health Sciences, Edith Cowan University, Perth, Western Australia, Australia

6 Research Center for Biochemistry and Molecular Biology and Jiangsu Key Laboratory of Brain Disease Bioinformation, Xuzhou Medical University, Xuzhou, China

⁷Department of Laboratory Medicine, Affiliated Hospital of Xuzhou Medical University, Xuzhou, China

Correspondence

Su‐Hua Qi and Bing Gu, School of Medical Technology, Xuzhou Medical University, Tongshan Rd 209, 221004 Xuzhou, China. Email: suhuaqi@xzhmu.edu.cn and gubing@gdph.org.cn

Abstract

Depression is a common and debilitating condition for which effective treatments are needed. Lepidium meyenii Walp (Maca) is a plant with potential medicinal effects in treating depression. Recently, there has been growing interest in plant-derived extracellular vesicles (EVs) due to their low toxicity and ability to transport to human cells. Targeting the gut–brain axis, a novel strategy for depression management, may be achieved through the use of Maca-derived EVs (Maca-EVs). In this study, we successfully isolated Maca-EVs using gradient ultracentrifugation and characterized their shape, size, and markers (CD63 and TSG101). The in vivo imaging showed that the Dil-labeled Maca‐EVs crossed the brain–blood barrier and accumulated in the brain. The behavioral tests revealed that Maca-EVs dramatically recovered the depression-like behaviors of unpredictable chronic mild stress (UCMS) mice. UCMS mice fecal were characterized by an elevated abundance of g_Enterococcus, g_Lactobacillus, and g_Escherichia_Shigella, which were significantly restored by administration of Maca-EVs. The effects of Maca-EVs on the altered microbial and fecal metabolites in UCMS mice were mapped to biotin, pyrimidine, and amino acid (tyrosine, alanine, aspartate, and glutamate) metabolisms, which were closely associated with the serotonin (5‐HT) production. Maca‐EVs were able to increase serum monoamine neurotransmitter levels in UCMS mice, with 5‐HT showing the most significant changes. We further demonstrated that 5‐HT improved the expression of brain‐derived neurotrophic factor, a key regulator of neuronal plasticity, and its subsequent activation of TrkB/p‐AKT signaling by regulating the GTP‐Cdc42/ERK pathway. These findings suggest that Maca‐EVs enhance

Rui Hong, Lan Luo, Liang Wang, and Zhao‐Li Hu contributed equally.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. iMeta published by John Wiley & Sons Australia, Ltd on behalf of iMeta Science.

$\frac{2 \text{ of } 18}{2 \text{ of } 18}$ HONG ET AL.

Funding information

Young Science and Technology Innovation Team of Xuzhou Medical University, Grant/Award Numbers: TD202005, TD202001; Jiangsu Qing‐Lan Project, Grant/Award Number: Year 2020; Leadership program of Xuzhou Medical University,

Grant/Award Number: JBGS202203; Natural Science Foundation of Jiangsu Province, Grant/Award Number: BK20211348; Scientific Research Project of Jiangsu Provincial Healthy Commission, Grant/Award Number: ZDB2020024

5‐HT release, possibly by modulating the gut–brain axis, to improve depression behavior. Our study sheds light on a novel approach to depression treatment using plant‐derived EVs.

KEYWORDS

5-HT, brain-derived neurotrophic factor, depression, gut-brain axis, Lepidium meyenii Walp‐derived extracellular vesicles, unpredictable chronic mild stress

Highlights

- We successfully isolated Maca-EVs using ultracentrifugation and tested its antidepressant effects in the unpredictable chronic mild stress mice.
- Maca-EVs increased serum serotonin (5-HT) levels probably via the gut microbiota and metabolism modulation.
- The antidepressant activity of Maca-EVs may be attributed to the enhanced serum 5-HT levels and subsequent activation of the GTP-Cdc42/ERK and BDNF/TrkB/AKT signaling pathway.

INTRODUCTION

Depression is a mental disorder that affects 280 million individuals globally, including 5.0% of adults and 5.7% of the elderly. Severe depression can lead to suicide, which is the fourth leading cause of death in populations between the ages of 15–29. Recent data released by the WHO have demonstrated that a 25% increase in the global prevalence of depression due to the spread of COVID‐19 [\[1](#page-16-0)]. It has been found that the dysfunction in excitatory synapses, microglia, neurotransmitters, neuroimmune, and neuroinflammatory actions may contribute to the development of depression $[2-4]$ $[2-4]$. Consequently, drugs that regulate neurotransmitters such as serotonin (5‐HT), norepinephrine (NE), dopamine (DA), and others are included in the clinical management of depression [[5\]](#page-16-2). Traditional antidepressants such as monoamine oxidase inhibitors, tricyclic, selective 5‐HT reuptake inhibitor, 5‐HT, and noradrenaline reuptake inhibitors have been shown to have particular antidepressant efficacy. However, their adverse effects such as sexual dysfunction, nausea/vomiting, weight changes, sleep disruption, and easy addiction could not be ignored [\[6\]](#page-16-3). Furthermore, almost 50% of patients with depression do not respond to antidepressant treatment [[7\]](#page-16-4). Thus, there is an unmet need for the development of safe and effective antidepressant.

Over the past decade, a growing body of evidence has highlighted that the gut–brain axis action is essential in the pathophysiological development of depression [[8\]](#page-17-0). This axis refers to the bidirectional interactions between the gut and brain, facilitated by neural (enteric, sympathetic, and vagus nerves), immune (inflammatory cytokines and cells), and chemical (microbiota metabolites and neurotransmitters) signals. In individuals with depression, the activity of the enteric and sympathetic nervous systems is elevated. Recent findings have shown that the depression‐like behaviors induced by lipopolysaccharide or "depression‐related" microbes injection in mice could be alleviated by subdiaphragmatic vagotomy, demonstrating the potential role of the vagus nerve in depression [\[9, 10\]](#page-17-1). Additionally, depression is characterized by elevated levels of peripheral proinflammatory cytokines or chemokines, such as tumor necrosis factor (TNF- α), interleukin-6 (IL-6), and IL-1 β , and activation of immune cells such as neuron cells in the brain, dendritic cells, and innate lymphoid cells in the gut, monocytes, and macrophages in the systemic circulation [\[11](#page-17-2)]. Furthermore, certain gut microbiota can produce specific metabolites (e.g., tryptophan metabolites, trimethylamine‐N‐oxide, and short‐chain fatty acids), neuroactive modulators (e.g., 5‐HT, aminobutyric acid, brain‐derived neurotrophic factor [BDNF], and glia‐ derived neurotrophic factor), and other factors that are secreted by intestinal cells or from bacteria decomposition $[12]$ $[12]$. Therefore, the gut-brain axis is emerging as a novel target for drug development to treat depression.

Lepidium meyenii Walp, also known as Maca, has been cultivated in the Andean region for at least 2000 years and successfully introduced in China since 2002, including Yunnan, Xinjiang, Jilin, and Tibet Province [\[13\]](#page-17-4). Maca is a traditional edible medicine plant renowned for its benefits in hormone balance, regulation of sexual dysfunction, and energizing effects [\[13](#page-17-4)]. Maca is rich in essential nutrients and bioactive components that vary depending on its color and type, resulting in diverse biological functions [[14](#page-17-5)]. The three most used and studied types are yellow, red, and black Maca. Recently, Maca and its extract have garnered significant attention due to its potential neuroprotective and antidepressant effects. Studies have shown that Maca macamide repairs the corticosterone‐induced hippocampal impairments through its anti-inflammatory, neurotrophic, and synaptic protection features [\[15](#page-17-6)]. Additionally, yellow, red, and black Maca have demonstrated antidepressant activity in ovariectomized mice [[16\]](#page-17-7). Furthermore, the petroleum ether extract of Maca exhibited antidepressant action by activating the noradrenergic and dopaminergic systems and inhibiting oxidative stress in the mouse brain [[17\]](#page-17-8). Nevertheless, the limited ability of Maca and its extract to cross the blood–brain barrier (BBB) remarkably hinders their potential as therapeutic agents. Plantderived extracellular vesicles (PDEVs), similar to animal‐derived extracellular vesicles (EVs), have been shown to participate in human cell-to-cell communication, exerting various biological functions, such as innate immunity and inflammation modulation [\[18\]](#page-17-9). The small size of EVs (<200 nm) enables them to cross the BBB freely, making them promise new candidates for translational applications [[19\]](#page-17-10).

In this study, we successfully isolated and purified EVs from Maca (Maca‐EVs) based on the minimal criterion of EVs characterizations. Behavioral experiments indicated the Maca‐EVs had remarkable antidepressant effects in the unpredictable chronic mild stress (UCMS) mice. The antidepressant activity of Maca‐EVs may be attributed to the enhancement of serum 5‐HT levels and subsequent activation of the BDNF/TrkB/AKT axis. Possible mechanisms for the increased serum 5‐HT levels include modulation of the gut–brain axis, among others.

RESULTS

Isolation and characterization of Maca‐EVs

Maca‐EVs were isolated from Maca juice using gradient ultracentrifugation as described in Figure [1A.](#page-5-0) Transmission electron microscopy imaging and nanoparticle tracking analysis revealed that Maca‐EVs were irregular‐shaped membrane-enclosed vesicles (Figure [1B\)](#page-5-0) with an average diameter of 134 nm (Figure [1C](#page-5-0)). Moreover, Maca-EVs expressed higher EVs markers CD63 and TSG101 than the Maca medium (Figure $1D$), indicating the successful isolation of EVs from Maca juice. Next, we investigated the in vivo uptake of Maca‐EVs in C57BL/6J mice using an

ANTIDEPRESSANT EFFECTS OF MACA‐EVS \blacksquare 3 of 18

IVIS Spectrum imaging system. Intravenously administrated Dil labeled Maca‐EVs (Maca‐EVs‐Dil) accumulated the mice brain with robust red fluorescence signals compared with the free‐Dil administrated mice (Figure [1E\)](#page-5-0). These mice were killed 12 h later after Maca-EVs-Dil or free-Dil administration, and brainfrozen sections were prepared. Immunofluorescence staining confirmed that Maca‐EVs could pass through the BBB and accumulate in the brain (Figure [1F](#page-5-0)).

Maca‐EVs reverse the depressive‐like behaviors in the UCMS mice

Next, we assessed the antidepressant-like effect of Maca-EVs in vivo. We applied daily UCMS in C57BL/6J mice for 8 weeks to establish the depression model (Figure [2A\)](#page-6-0). Stress‐free treated mice were included as controls. UCMS dramatically deceased the body weight (Figure S1A), increased immobility time in the tail suspension test (Figure S1B) and forced swim test (Figure S1C), increased latency to groom and eat (Figure S1D,E), and decreased sucrose consumption (Figure S1F) in the mice. There were no significant changes in the time of open arm and close arm, indicating a depression model, but not an anxiety model (Figure S1G,H). Control mice were randomly divided into two groups: control and control injected with Maca‐EVs (100 μg/kg). UCMS mice were randomly divided into four groups: UCMS and UCMS injected with different doses of Maca-EVs (50, 100, and $200 \mu g/kg$). Maca‐EVs were administrated once every 2 days for a total of eight times in 2 weeks. Notably, we continuously applied daily UCMS to the mice during the Maca‐EVs treatment. Fecal and blood samples were collected from the mice, and the mice were killed after the behavioral tests were completed (Figure [2A](#page-6-0)). Body weights of the mice were continually declined in UCMS mice, but were comparable to the control mice administrated with Maca-EVs $(200 \mu g/kg)$ (Figure [2B\)](#page-6-0). Maca-EVs administration improved depressive‐like behaviors in UCMS mice, as shown by the decline in the extended immobility time in the tail suspension test (Figure $2C$) and the forced swim test (Figure [2D](#page-6-0)). The open field test (Figure [2E\)](#page-6-0) showed that Maca‐EVs administration improved exploratory behavior with elevated traveling distances (Figure [2F](#page-6-0)) and entry frequency in the zone (Figure [2G](#page-6-0)) of UCMS mice. The sucrose consumption test demonstrated a significant reduction in the latency to eat (Figure [2H\)](#page-6-0) or groom (Figure [2I](#page-6-0)) and an increase in sucrose preference (Figure [2J\)](#page-6-0) in Maca‐EVs ‐administrated UCMS mice compared with the nontreated UCMS mice. These data suggested the therapeutic potential of Maca‐EVs in treating depression.

FIGURE 1 Isolation and characterization of Maca-EVs. (A) Flowchart on the isolation of EVs from Lepidium meyenii Walp (Maca). (B) Transmission electron microscope image on the structure of Maca‐EVs (red arrow). Scale bar: 200 nm. (C) NTA analysis revealed that the size of Maca‐EVs was around 134 nm. (D) Western blot analysis on the expressions of EVs markers (CD63 and TSG101) in the Maca medium and Maca-EVs. (E) Representative images on the in vivo location of Free-Dil or Maca-EVs-Dil in the brain of mice by the IVIS optical imaging system. (F) Representative immunofluorescence images on the brain tissue of mice that killed 12 h later after the injection of Free‐Dil or Maca‐EVs‐Dil. Scale bar: 100 μm. DAPI, 4′,6‐diamidino‐2‐phenylindole; EV, extracellular vesicle; IVIS, in vivo imaging system; NTA, nanoparticle tracking analysis; PBS, phosphate-buffered saline.

ANTIDEPRESSANT EFFECTS OF MACA‐EVS \blacksquare

Changes in the fecal microbiota in Maca‐EVs treated or nontreated control and UCMS mice

We next investigated the dynamic changes of gut microbiota in mice from across four groups (Control,

Control + Maca‐EVs, UCMS, and UCMS + Maca‐EVs), using 12 fecal samples. A total of 718,971 high-quality 16S ribosomal RNA (rRNA) sequences were obtained with an average of 59,914 reads per sample. After the rarefaction of sample to an equal sequencing depth (56,402 reads per sample) and clustering, 676,824

$\frac{6 \text{ of } 18 \text{ }}{4 \text{ N}}$ WILEY-**iMeta**

sequences were grouped into 1434 operational taxonomic units (OTUs) for the downstream analysis. At the phylum level, Bacteroidota and Firmicutes (Figure [3A\)](#page-8-0) were the most abundant in all groups, while at the genus level, Muribaculaceae was the most abundant in all groups (Figure [3B](#page-8-0)). The genus Lactobacillus varied among groups, with a significant decrease in Control, Control + Maca-EVs, and UCMS + Maca‐EVs groups compared with UCMS group ($p < 0.05$, Figure [3B\)](#page-8-0). In contrast, the abundance of Akkermansia, Dubosiella, and Faecalibaculum was significantly lower in the UCMS group than that in the other groups (Figure [3B](#page-8-0)). These data indicated that the genera Lactobacillus, Akkermansia, Dubosiella, and Faecalibaculum might be associated with the effect of UCMS and Maca‐EVs.

We also investigated the effect of Maca‐EVs on microbial diversity and found that microbial richness, as indicated by the Faith‐pd and Observed features index, was higher in the Maca-EVs groups (Control + Maca-EVs and UCMS + Maca‐EVs), compared with Control and UCMS groups (Figure $3C$,D). Only the Faith-pd index in Control + Maca‐EVs group was significantly lower than that in the Control group $(p < 0.05$, Figure [3D\)](#page-8-0). The community richness of the UCMS group relative to the Control group decreased but not significantly. To evaluate the changes of β -diversity of gut microbiota across different groups, we conducted a principal coordinate analysis based on the Bray‐curtis distance matrixes (Figure $3E$) and partial least squaresdiscriminant analysis (PLS‐DA) (Figure S2B). We found that the four groups were clustered into distinct groups with β-diversity estimates, as assessed by the ANOSIM tests $(R = 0.772, p = 0.001)$.

To obtain a deeper insight of microbiota alterations upon Maca‐EVs administration, we performed linear discriminant analysis effect size (LEfSe) to compare microbial composition at 16S recombinant DNA OTU level. The Venn diagram (Figure S2A) showed

that 444 OTUs were shared between Control and Control + Maca‐EVs groups, while 494 were unique to Control group and 222 to Control + Maca-EVs group. Between Control and UCMS groups, 578 OTUs were shared, while 360 were unique to Control group and 375 to UCMS group. Between UCMS and UCMS + Maca‐EVs groups, 471 OTUs were shared, while 382 were unique to UCMS group and 209 to UCMS + Maca‐EVs group. We grouped Control, Control + Maca‐EVs, and UCMS + Maca‐EVs into a new group called New_Control. A total of 11 OTUs that significantly differed between New_Control and UCMS groups ($p < 0.05$; linear discriminant analysis >4.0) were shown in Figure [3F](#page-8-0), of which 1 OTU was enriched in New_Control group and 10 in the UCMS group. Among those differential OTUs, 2 OTUs belong to order Lactobacillales, which are Lactobacillaceae and Enterococcaceae at the family level. To further investigate which OTUs were associated with depressivelike symptoms, we selected four typical bacteria (Figure [3G](#page-8-0)–J) that were significantly differed between New_Control and UCMS groups to see their abundance changes among the four groups. Furthermore, we selected four typical bacteria that were significantly different between the Control and UCMS groups (Figure S2C–F).

Changes in the fecal metabolic signatures of Maca‐EVs treated or nontreated control and UCMS mice

Given gut microbiome often modulates the host metabolic pathway, here we used gas chromatography–mass spectrometry (GC-MS)-based metabolomics to compare the metabolic characteristics of the four groups. To detect more metabolites, nontargeted metabolomics was performed in two modes, positive ion (pos) and negative ion (neg). Principal components analysis and PLS‐DA were conducted to evaluate the changes of fecal metabolic

FIGURE 2 Antidepressive effects of Maca-EVs in the UCMS mice. (A) Schematic illustration on the UCMS model establishment and drug administration in the present study. (B) Body weight changes of control and UCMS mice with indicated Maca-EVs treatments at different time points. Effect of Maca‐EVs on the immobility time in tail suspension test (C) and in forced swimming test (D) of control and UCMS mice with indicated Maca‐EVs treatments. (E) Representative open field test photos of control and UCMS mice with indicated Maca-EVs treatments. Quantification on the total traveling distance (F) and entries in zone (G) of the open field test. Effect of Maca-EVs on the latency to eat (H) and to groom (I), and percentage of sucrose preference (J) of control and UCMS mice with indicated Maca-EVs treatments. All data are presented as mean \pm SEM ($n = 15-30$ experiments for each group). Significance was evaluated by RM two-way analysis of variance (ANOVA) followed by Tukey's multiple comparisons tests in (B), and significance was evaluated by ordinary one‐way ANOVA followed by Tukey's multiple comparisons test in (C, F, G), Brown–Forsythe and Welch ANOVA tests followed by the Tamhane T2 multiple comparisons test in (D), and the Kruskal–Wallis test followed by Dunn's multiple comparisons test in (H, I, J). Means denoted by a different letter indicated significant differences between groups ($p < 0.05$). EV, extracellular vesicle; RM, repeated measure; SEM, scanning electron microscope; UCMS, unpredictable chronic mild stress.

FIGURE 3 Effects of Maca-EVs on the microbiota composition in Control and UCMS mice feces. Mice were killed when the behavior tests were accomplished, and feces were collected for 16S rRNA sequencing analysis. Control, Control + Maca-EVs, UCMS, and UCMS + Maca-EVs groups were included in the present study ($n = 3$ experiments for each group). Relative abundance of microbiota (top 10) at phylum (A) and genus (B) levels in the different groups. Alpha diversity regarding the index of Faith's phylogenetic diversity metric (pd) (C) and observed features (D) in the different groups. (E) Principal coordinate analysis (PCoA) based on the Bray-curtis distance matrixes between sample groups. (F) Linear discriminant analysis effect size (LEfSe) was performed to compare microbial composition at 16S rDNA operational taxonomic unit (OTU) level between sample groups. A total of 11 OTUs that significantly differed between New_Control and UCMS groups ($p < 0.05$; linear discriminant analysis [LDA] > 4.0) were shown. (G–J) The abundance changes of typical bacteria that were significantly differed between New_Control and UCMS among sample groups. EV, extracellular vesicle; rDNA, recombinant DNA; rRNA, ribosomal RNA; UCMS, unpredictable chronic mild stress.

signatures across different groups (Figure S3A–D). Specifically, Figure S3A,S3B was obtained from pos mode, Figure S3C,S3D was obtained from neg mode. Overall, the results showed spatial separation of metabolic signatures among the four groups, particularly between New_Control group and UCMS group, indicating compositional differences between them. In pos mode, 63 differential metabolites were identified between

FIGURE 4 Effects of Maca-EVs on the metabolomics in control and UCMS mice feces. (A) Differential metabolites between New_Control and UCMS group (VIP > 1, $p < 0.05$, $\log FC$) = 1) in pos mode. (B–E) The levels of four typical metabolites that were significantly differed between New_Control and UCMS groups among sample groups. (F) Reciprocal interactions between altered gut bacteria and serum metabolites identified by a co‐occurrence network based on Spearman correlation analysis in pos mode. EV, extracellular vesicle; UCMS, unpredictable chronic mild stress. VIP, variable importance in projection.

New_Control and UCMS group (variable importance in projection $[VIP] > 1$, $p < 0.05$, $\log FC > 1$) (Figure [4A\)](#page-9-0). These included Vanillin, L-5-hydroxytryptophan, 4-(2,3dihydro‐1,4‐benzodioxin‐6‐yl) butanoic acid, 6‐methoxy‐ 2‐naphthoic acid enriched in New_Control group, and Prostaglandin I2, Lysops 22:5, N‐acetylornithine, N‐acetyl‐ aspartic acid enriched in UCMS group. In neg mode (Figure S3G), 47 differential metabolites were found between New_Control and UCMS group $(VIP > 1$, $p < 0.05$, $\log FC$ > 1). Metabolites enriched in New_Control group included N‐isobutyrylglycine, N‐acetylalanine, pantothenic acid, 2‐keto‐4‐methylthiobutyric acid, and so forth, while metabolites enriched in the UCMS group included lysophosphatidylethanolamine 18:1, Apigenin, Saccharin, 5‐methoxysalicylic acid, and so forth. We selected four typical metabolites (Vanillin, ^L‐ 5‐hydroxytryptophan [5‐HTP], 4‐(2,3‐dihydro‐1,4‐ benzodioxin‐6‐yl) butanoic acid, and 6‐methoxy‐2‐ naphthoic acid) that were significantly differed between the New_Control and UCMS groups and analyzed their changes among the four groups (Figure [4B](#page-9-0)–E). Pathway enrichment analysis indicated that UCMS caused major alterations in metabolic pathway related to biotin, pyrimidine, tyrosine, alanine, aspartate, and glutamate metabolism (Figure S3E). By tryptophan hydroxylase converts tryptophan into 5‐hydroxytryptophan (5‐HTP), which is then further converted to 5‐HT by 5‐HTP decarboxylase [\[20](#page-17-11)] (Figure S3F). In our metabolomics analysis, we found that the amount of 5‐HTP in this pathway was significantly lower in the UCMS group but higher in the UCMS + Maca-EVs group (Figure $4C$), suggesting that UCMS may affect 5‐HT biosynthesis. Taken together, these results suggest that metabolites are associated with distinct groups.

Co‐occurrence network analysis between the gut microbiota and fecal metabolites

To investigate the potential reciprocal interactions between altered gut bacteria and metabolites, a cooccurrence network was constructed based on Spearman correlation analysis (Figure [4F](#page-9-0)). In pos mode, we found that the genus Lactobacillus formed a strong co‐occurring relationship with fecal metabolites assigned to 2‐piperidinobenzoic acid, R‐1 methanandamide phosphate, polyphosphate kinase (PPK), isohomovanillic acid, and N6‐acetyl‐L‐lysine. The genus Enterococcus formed a strong co‐occurring relationship with fecal metabolites assigned to lysophosphatidylcholine (LPC) 15:0, D‐(‐)‐Salicin, ACar 16:1, 4‐Pregnen‐ 17,20‐Diol‐3‐One, leucine (L)‐asparagine (N)‐lysine (K) (LNK), N6‐acetyl‐L‐lysine. The genus Escherichia_Shigella

ANTIDEPRESSANT EFFECTS OF MACA‐EVS \blacksquare

formed a strong co-occurring relationship with fecal metabolites assigned to pregnenolone, delta-tridecalactone, estropipate, 2‐deoxyuridine, and deoxycytidine. The genera Lactobacillus was positively correlated with PPK, N6‐acetyl‐ ^L‐lysine, negatively correlated with 2‐piperidinobenzoic acid, R‐1 methanandamide phosphate, and isohomovanillic acid. The genus Enterococcus was positively correlated with 4‐ Pregnen‐17,20‐Diol‐3‐One, LNK, and N6‐acetyl‐L‐lysine, negatively correlated with LPC 15:0, D‐(‐)‐Salicin and ACar 16:1. The genus Escherichia_Shigella was positively correlated with 2‐deoxyuridine and deoxycytidine, negatively correlated with pregnenolone, delta-tridecalactone, and estropipate. Taken together, the altered Enterococcus, Lactobacillus, and Escherichia_Shigella were mapped to biotin and pyrimidine metabolism in pos mode. We can also see co-occurrence network in neg mode which showed that the altered Enterococcus, Lactobacillus, and Escherichia_Shigella were mapped to tyrosine, alanine, aspartate, and glutamate metabolism (Figure S3F,G). Notably, these findings indicate that altered gut microbiota and metabolites formed a synergistic and node-related cooccurrence network between the New_Control and UCMS groups.

Maca‐EVs enhance serum 5‐HT expressions in the UCMS mice

As the identified altered metabolisms were closely associated with 5‐HT production, we next investigated whether Maca-EVs modulate the monoamine neurotransmitters production. We collected the mice serum after completing the behavioral tests to measure the levels of the three major monoamine neurotransmitters (5‐HT, NE, and DA) via HPLC. The signals of NE and DA were too low to quantify. We observed significantly downregulated serum 5‐HT levels in the UCMS mice compared with the control mice, confirming the successful establishment of the UCMS model (Figure [5A,B\)](#page-11-0). Maca‐EVs administration dramatically increased serum 5‐HT levels in the UCMS model at all doses. The significantly elevated serum NE and DA levels in the UCMS mice were observed when treated with Maca‐EVs that started from 100 to 200 μg/kg doses. While the serum inflammatory cytokines IL‐1β, IL‐6, and TNF‐α levels were increased in the UCMS mice compared with the control mice, the differences were not significant (Figure $5C-E$ $5C-E$). At a dose of 200 μg/kg, Maca-EVs significantly reduced TNF- α levels in serum, while the decrease in IL‐1β and IL‐6 expression was not statistically significant. Collectively, the serum monoamine neurotransmitters levels, particularly 5‐HT, may contribute to the antidepressant effects of Maca-EVs.

FIGURE 5 Serum monoamine neurotransmitters and inflammatory cytokines expressions analyses. (A) The UPLC‐MS/MS chromatograms of monoamine factors, including 5‐HT in the standard and serum of control and UCMS mice with indicated Maca‐EVs treatments. (B) Quantitative data on the concentrations of 5‐HT in the serum of control and UCMS mice with indicated Maca‐EVs treatments. Quantitative data on the concentrations of inflammatory cytokines, including IL-1β (C), IL-6 (D), and TNF- α (E) in the serum of control and UCMS mice with indicated Maca-EVs treatments. All data are presented as mean \pm SEM ($n = 3-11$ experiments for each group). Significance was evaluated by ordinary one‐way analysis of variance (ANOVA) followed by Tukey's multiple comparisons test in (B, C, E) and the Kruskal–Wallis test followed by Dunn's multiple comparisons test in (D). Means denoted by a different letter indicated significant differences between groups ($p < 0.05$). 5-HT, serotonin; EV, extracellular vesicle; IL, interleukin; MS, mass spectrometry; SEM, scanning electron microscope; UCMS, unpredictable chronic mild stress; UPLC, ultraperformance liquid chromatography.

Maca‐EVs attenuate the decrease in the BDNF expression by activating the GTP‐Cdc42/ERK signaling pathway in the hippocampus and cortex of UCMS mice

The coregulation of 5‐HT and BDNF signaling is well established, whereby 5‐HT activates the BDNF expression, which is essential for the growth and survival of 5‐HT neurons. Abnormal 5‐HT and BDNF signaling are associated with the progression of depression $[21]$ $[21]$. The administration of Maca-EVs at a dose of 200 μg/kg significantly increased serum 5‐HT levels, promoting us to investigate whether it would enhance BDNF expression in the UCMS mice. Using the immunofluorescence staining and WB analysis, we confirmed the downregulation of BDNF expression in the hippocampus (Figure [6A,B\)](#page-12-0) and cortex (Figure S4A,B) of the UCMS mice, which was attenuated by Maca‐EVs administration. To further explore the underlying mechanism, we examined the GTP‐Cdc42/ERK signaling pathway that regulates BDNF expression. The WB analysis revealed an upregulated ratio of GTP‐Cdc42/Cdc42 and p‐ERK/ ERK in the hippocampus (Figure $6C$) and cortex (Figure S4C) of the Maca‐EVs administrated UCMS

mice compared with the saline‐administrated UCMS mice. These results suggest that Maca‐EVs attenuate the decrease in BDNF expression by triggering the GTP‐Cdc42/ERK signaling in the hippocampus and cortex of UCMS mice.

Maca‐EVs mitigate the neuron number decline via modulating the TrkB/AKT signaling in the hippocampus and cortex of UCMS mice

Moreover, we explored whether the upregulated BDNF expression modulates neuron growth in UCMS mice. BDNF interacts with its receptor Tyrosine Kinase receptor B (TrkB) and initiates the downstream AKT signaling to maintain neuron growth. We observed downregulated TrkB expression and upregulated p‐AKT/AKT ratio in the UCMS mice, which were significantly rescued by Maca-EVs administration (Figures [6D](#page-12-0) and S4D,E). The immunofluorescence staining revealed that the Maca‐EVs administration promoted neuron growth in the dentate gyrus zone of UCMS mice, which was declined (Figures [6E](#page-12-0) and S4F). Therefore, the data demonstrated that Maca‐EVs mitigated the declined number of neurons

FIGURE 6 (See caption on next page)

 $\frac{12 \text{ of } 18 \text{ }}{1000 \text{ F}}$ HONG ET AL.

The 5‐HT induces BDNF expression and subsequent activation of TrkB/AKT signaling via modulating the GTP‐Cdc42/ ERK pathway in vitro

To determine whether the 5‐HT activation regulates the activity of GTP‐Cdc42, we treated PC12 cells with ML141, a GTP‐Cdc42 inhibitor, and observed a significant decrease in the ratio of GTP‐Cdc42/Cdc42 and p‐ERK/ERK (Figure [6F](#page-12-0)). We also found that the upregulation of BDNF expression induced by 5‐HT was mitigated by the ML141 treatment (Figure $6G$, H), indicating that 5‐HT stimulates BDNF expression through modulation of the GTP‐Cdc42/ERK pathway. Moreover, ML141 treatment blocked the subsequent TrkB/AKT signaling of 5‐HT, as evidenced by the declined TrkB expression and ratio of p-AKT/AKT (Figure [6I,J\)](#page-12-0). These data suggest that 5‐HT activates the GTP‐Cdc42/ERK pathway to stimulate BDNF expression and subsequent activation of TrkB/AKT signaling.

DISCUSSION

To the best of our knowledge, we are the first to successfully isolate and characterize EVs from Maca. Maca-EVs exhibited features similar to animal cellderived EVs and could pass through the BBB. Intravenous injection of Maca‐EVs noticeably reduced the depressive behaviors in UCMS mice. Our analysis, using 16S rRNA and GC‐MS‐based metabolomics, identified that Maca‐EVs altered the composition of depression‐ related fecal microbiota and improved metabolisms associated with 5‐HT production in UCMS mice. This suggests the potential of Maca‐EVs to modulate the gut–brain axis. Furthermore, we found that Maca‐EVs significantly enhanced serum 5‐HT levels, which were decreased in UCMS mice, as determined by HPLC analysis. In vivo and in vitro experiments revealed that 5‐HT stimulates BDNF expression and consequent activation of BDNF/TrkB/AKT signaling, which may contribute to the antidepressant effects of Maca‐EVs.

Traditional Chinese Medicine (TCM) has long been widely used in the treatment of depression, particularly in Asia. The molecular mechanisms underlying the action of TCM involve the regulation of monoamine transmission, hypothalamic‐pituitary‐adrenal axis, neurotrophins, such as BDNF, as well as the number and function of synapses [\[22\]](#page-17-13). Maca extract has been reported to possess neuroprotective and antidepressant effects attributed to its antioxidant activity [[23\]](#page-17-14). Later studies identified macamides as the active constituents responsible for the neuroprotection by inhibiting fatty acid amide hydrolase, which regulates neural progenitor cell proliferation [[24\]](#page-17-15). In addition, Maca extract has been suggested to have anti-inflammatory properties by reducing the inflammatory cytokine IL‐6 in Maca users [\[25](#page-17-16)]. However, the precise molecular mechanisms of Maca and its extract in treating depression remain to be elucidated.

Over the past decades, PDEVs, which are spherical‐, oval‐, or cup‐shaped vesicles containing biologically active elements, including lipids, proteins, nucleic acids, and secondary metabolites, have gained substantial attention in therapeutic drug development. Increasing in vitro and in vivo studies have shown that PDEVs possess anticancer, anti-inflammatory, antioxidant, and regenerative activities [[26\]](#page-17-17). Of note, Teng et al. identified that microRNA (miRNA) from ginger‐derived PDEVs can target the gut bacteria Lactobacillus rhamnosus genome to produce more indole‐3‐carboxaldehyde, which promotes IL‐22 secretion, enhancing gut barrier

FIGURE 6 Maca-EVs modulated the BDNF/GTP-Cdc42/ERK and TrkB/AKT signaling in the hippocampus of mice via the 5-HT. (A) Representative immunofluorescence images on the BDNF expressions in the hippocampus of control, Maca-EVs treated (200 μg/kg), or untreated UCMS mice. Scale bar: 50 μm. Western blot images and quantitative data on the expressions of BDNF (B), Cdc42 enzyme activity, p‐ERK and ERK (C), TrkB, p‐AKT and AKT (D) in the hippocampus of control, Maca‐EVs untreated or treated (200 μg/kg) UCMS mice. (E) Representative immunofluorescence images on the NeuN cells in the hippocampus of control, Maca‐EVs treated (200 μg/kg), or untreated UCMS mice. The yellow dashed box indicated the dentate gyrus zone of hippocampus. Scale bar: 50 μm. (F) PC12 cells were treated with 5‐HT or ML141 (a GTP‐Cdc42 inhibitor) for 12 h and cultured for another 24 h. Western blot images and quantitative data on the expressions of GTP‐Cdc42, Cdc42, p‐ERK, and ERK in PC12 cells were shown. Immunofluorescence (G) and western blot analysis (H) and on the BDNF expressions in PC12 cells. Scale bar: $50 \mu m$. Western blot images and quantitative data on the expressions of TrkB (I), p-AKT, and AKT (J) in PC12 cells. All data are presented as mean \pm SEM ($n = 3$ experiments for each group). Significance was evaluated by ordinary one‐way analysis of variance (ANOVA) followed by Tukey's multiple comparisons test in (B, C, D, F, H, I, J). ns, not significant, $p > 0.05$, * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$ between two indicated groups. 5-HT, serotonin; BDNF, brain-derived neurotrophic factor; DAPI, 4′,6‐diamidino‐2‐phenylindole; EV, extracellular vesicle; SEM, scanning electron microscope; UCMS, unpredictable chronic mild stress.

function [[27\]](#page-17-18). Despite promising preclinical studies, the translation of PDEVs from bench to bedside is still in its infancy. At present, the University of Louisville has conducted three clinical trials (NCT04879810, NCT01668849, and NCT01294072) using four types of PDEVs (curcumin, ginger, aloe, and grape) [[28\]](#page-17-19). However, the complete results of these clinical trials have yet to be reported. Therefore, significant efforts are needed to focus on the isolation, purification, physiochemical characterization, quality control, safety, and delivery routes of PDEVs [[29\]](#page-17-20).

Our group has previously isolated EVs from Momordica charantia, a fruit of the Cucurbitaceae plant, and demonstrated its therapeutic potential in treating cerebral ischemia/reperfusion $\begin{bmatrix} 30 \end{bmatrix}$ and radiation-induced heart disease [\[31](#page-18-1)]. Hence, we aimed to isolate EVs from black Maca and investigate their antidepressant potency in a UCMS mice model, along with the relevant mechanisms. We successfully isolated Maca-EVs by gradient centrifugation, which exhibited features similar to animal cell-derived EVs (Figure [1A](#page-5-0)–D). We used the lipophilic fluorescent dye Dil, which is widely used for labeling EVs, to characterize and monitor the cellular uptake or tissue distribution of Mca‐EVs. The in vivo imaging showed that Dil‐Maca‐EVs efficiently crossed the BBB and had a higher fluorescent density in the mouse brain (Figure $1E, F$). Moreover, we demonstrated the efficacy of Maca‐EVs in treating depressed UCMS mice using behavioral tests, such as tail suspension test, forced swim test, open field test, and sucrose consumption test, except for the regulation of sucrose preference (Figure [2](#page-6-0)).

The gut–brain axis represents bidirectional communication between the central nervous and digestive systems, involving neurotransmitters, neuroimmune, neuroendocrine, and sensory neural pathway [[32\]](#page-18-2). A good deal of data supported the role of the gut–brain axis link in the disease setting of depression $\left[33\right]$ $\left[33\right]$ $\left[33\right]$. Transplantation of depressed patients' fecal microbiota‐induced depressive‐like behaviors and physiological features in microbiota‐depleted rats, suggesting a causal role of gut microbiota in the development of depression [[34\]](#page-18-4). Additionally, increasing efforts have been devoted to depicting the characteristics of microbiota composition and metabolism in depressive patients. Raes et al. found that butyrate‐producing Faecalibacterium and Coprococcus bacteria were absent in depression patients [\[35](#page-18-5)]. In this study, we found that genus Lactobacillus showed a higher abundance in UCMS mice, while the abundance of Akkermansia, Dubosiella, and Faecalibaculum was higher in Control mice than UCMS mice, suggesting their potential role in the antidepressive effects of Maca-EVs (Figure $3A,B$). In addition, the α -diversity and

ANTIDEPRESSANT EFFECTS OF MACA‐EVS \blacksquare

β‐diversity of the microbiota were varied in the four groups (Figure [3C](#page-8-0)–E). By regrouping Control, Control + Maca‐EVs, and UCMS + Maca‐EVs into a new group (New_Control), we further demonstrated that Maca‐EVs administration reduced the abundance of g_Enterococcus and g_Lactobacillus, which were increased in UCMS mice (Figure [3G,H](#page-8-0)). Most human gut microbiota analysis identified that the abundance of g_Enterococcus and g_Lactobacillus were depleted in depressive patients [\[36\]](#page-18-6). Nevertheless, their abundance in depressed mice failed to reach an agreement. The abundance of g_E terococcus and g_Lactobacillus was found to increase in Bharwani's and study [\[37\]](#page-18-7), but decrease in the studies of Farshim et al. [\[38](#page-18-8)] and Galley et al. [\[39\]](#page-18-9). It must be admitted that there were inconsistences in the abundance, α‐diversity, and β‐diversity of gut microbiota composition due to individual and species differences [\[36\]](#page-18-6). Yang et al. described notably elevated g_Bacteroides and declined g_Blautia and g_Eubacterium with altered γ‐aminobutyrate, phenylalanine, and tryptophan metabolism in patients with the major depressive disorder [\[40\]](#page-18-10). Here, the Pathway enrichment analysis indicated that UCMS major alterations in metabolic pathway were mapped to biotin, pyrimidine, tyrosine, alanine, aspartate, and glutamate metabolism (Figures [4F](#page-9-0) and S3E). Moreover, the level of Tyrosine metabolism‐related product ^L‐5‐hydroxytryptophan was declined in UCMS mice but increased by Maca‐EVs administration (Figure [4C](#page-9-0)). These suggest that Maca‐EVs increase serum 5‐HT levels through modulation of gut microbiota composition and metabolism.

Excitatory or inhibitory neurotransmitters play an essential role in the homeostatic regulation of neuronal excitability, conferring normal brain functions. The theory that lowered excitatory neurotransmitter 5‐HT levels or activity are associated with depression remains influential. Recently, Moncrieff et al. implied that reduced 5‐HT levels or activity might not be the cause of depression [[41\]](#page-18-11). Nevertheless, currently prescribed drugs for treating depression mainly function by elevating excitatory neurotransmitters 5‐HT, NE, and DA levels throughout the body. Rizzo et al. demonstrated that selective serotonin reuptake inhibitor fluoxetine enhanced 5‐HT expressions, promoting neural stem cell proliferation, which was reversed by 5‐HT receptor antagonism $[42]$ $[42]$. In our study, all doses of Maca-EVs significantly increased serum 5‐HT levels in the UCMS mice (Figure [5B\)](#page-11-0). Innate and adaptive immune dysfunctions have been demonstrated to be involved in the pathophysiology of depression [\[43\]](#page-18-13). Grosse et al. determined that monocyte gene expression was age related in major depressive disorder patients [[44\]](#page-18-14). Thus, we investigated whether Maca‐EVs would modulate the

$\frac{14 \text{ of } 18}{1000 \text{ F}}$ HONG ET AL.

inflammatory responses by examining the three specific serum inflammatory cytokines, IL‐1β, IL‐6, and TNF‐α, which have been proved to be positively associated with depression [\[45](#page-18-15)]. In our model, increased serum IL‐1β, IL‐ 6, and TNF- α were detected in the UCMS mice, but were not statistically significant (Figure [5C](#page-11-0)–E). This may be because we tested the inflammatory factors at a late phase of depression induction. Nevertheless, Maca‐EVs significantly decreased the IL‐6 and TNF‐α levels only at a dose of 200 μg/kg. These data suggested that the 5‐HT signaling is more sensitive to the Maca-EVs treatment, and we next focused on its action in treating depressive UCMS mice in this study.

There are synergistic effects between 5‐HT and BDNF in modulating synaptic plasticity that 5‐HT triggers the expressions of BDNF, and BDNF improves 5‐HT neuronal growth and survival and maintains synaptic plasticity in the adult brain $[21]$ $[21]$. The activation of 5-HT signaling requires the interaction with 5‐HT receptors that is not only coupled with but also activates the G‐protein (Rho, Rac, and Cdc42)-related pathway [\[46](#page-18-16)]. Udo et al. demonstrated that the 5‐HT activated GTP‐Cdc42 in aplysia sensory neurons, promoting learning‐related synaptic growth $[47]$ $[47]$. Elena et al. identified that the activation of the 5‐HT receptor leads to filopodia formation via a Cdc42‐mediated pathway accompanied with RhoA relevant cell rounding in neuroblastoma cells [\[48](#page-18-18)]. Errico et al. [[49\]](#page-18-19) and Speranza et al. [[50](#page-19-0)] showed that the 5‐HT signaling activation enhanced neuron function via the ERK pathway. Others also identified that (R)‐ketamine isomer exerts antidepressant effects via activating the ERK‐NRBP1‐CREB‐BDNF pathway [[51,](#page-19-1) [52\]](#page-19-1). Indeed, elevated ratios of GTP‐Cdc42/Cdc42 and p‐ ERK/ERK, and BDNF expression were noted in the cortex and hippocampus of Maca‐EVs‐injected UCMS mice (Figures [6A](#page-12-0)–C and S4A–C). Evidence has suggested that BDNF signaling via its receptor TrkB is demanded for the effects of antidepressant drugs. Adachi et al. found that the absence of TrkB but not the deletion of BDNF caused decreased capacity in treating depression [\[53](#page-19-2)]. He et al. showed that the Alpinia oxyphylla Miq. Hippocampal neurogenesis was promoted via activating BDNF/TrkB/AKT signaling pathway [[54\]](#page-19-3). Accordingly, we also found that the Maca‐EVs increased the expression of TrkB and phosphorylated AKT in the cortex and hippocampus of UCMS mice (Figures [6D](#page-12-0) and S4D, E). The in vitro assay further confirmed that 5‐HT enhanced BDNF expression and consequent stimulation of TrkB/AKT signaling via the GTP‐Cdc42/ERK pathway (Figure [6F](#page-12-0)–J).

Unfortunately, we only initially isolated Maca‐EVs and investigated the antidepressant effect. We observed that Maca‐EVs could influence the fecal

microbiota composition and elevate serum 5‐HT levels, leading to increased BDNF expression via activation of the GTP‐Cdc42/ERK signaling pathway. Enterochromaffin cells of the intestinal mucosa are the primary site for the 5‐HT synthesis, storage, and release. We revealed a potential role of Maca‐EVs in modulating fecal microbiota/metabolite to produce 5‐HT, which requires further experiments to clarify. However, the EVs could cross the BBB and they may exert their antidepressant therapeutic activities by directly affecting the brain or through the peripheral pathway. In this study, we only focused on the effects of Maca‐EVs on modulating the gut–brain axis. The administration of Maca‐EVs via gavage or targeted delivery to the brain would be employed in future studies to elucidate the underlying mechanisms of Maca‐EVs in treating depression.

There are several other limitations in the present study. First, Maca‐EVs produce antidepressant‐like effects similar to those of Maca or its extract, possibly because Maca‐EVs contain them. Notably, the nanoscale size of Maca-EVs (<200 nm) allows them to overcome the disadvantages of Maca extract components, like, polysaccharides, proteins, and alkaloids (which are high molecular and insoluble), thereby enabling them to cross the BBB or intestinal barrier. Nevertheless, thorough omics studies should be performed to understand the biologically active constituents, such as nucleic acids, proteins, lipids, and secondary metabolites, to validate this speculation. Besides, the key elements that confer the antidepressant mechanism of Maca‐EVs remain unexplored. Recently, EVs containing miRNAs have attracted enormous interest in commercial spans as next-generation medicine [\[55](#page-19-4)]. However, a single miRNA may not be as effective as multiple miRNAs. Occasionally, even though some miRNAs reveal excellent therapeutic capacity, it is challenging to develop into drugs due to their ease of degradation, multiple targets, and the potential biological safety risk. For instance, the Phase 1 clinical trial (NCT02862145) on evaluating MRX34's (miRNA‐34) efficacy in treating melanoma was withdrawn due to severe immune-related side events. Emerging studies pointed out that the EVs' therapeutic activity originated from the synergetic effects of their contents $[56]$ $[56]$. Thus, it appears to be less important to identify the precise element that plays an essential role in the function of EVs. Second, we only unraveled the role of 5‐HT/BDNF in the antidepressant activity of Maca‐EVs. Apart from the BDNF, other neurotrophic factors including vascular endothelial growth factor, fibroblast growth factor 2, insulin‐like growth factor 1, and Activin‐A are also of interest to study. The mechanisms involved in two other elevated

monoamine neurotransmitters in antioxidant or anti-inflammatory effects require further investigation. Lastly, a sucrose test that may affect the composition of intestinal flora was conducted to assess the depression behavior, though our data showed the Maca‐EVs did not influence the sucrose preference level. In the future study, we will avoid these technical points in investigating the exact mechanism of Maca‐EVs in modulating the gut‐grain axis and their associations with Maca‐EVs' therapeutics in treating depression.

CONCLUSION

Taken together, this study reports for the first time the successful isolation and characterization of Maca-EVs, which exhibit antidepressant effects in UCMS mice. These effects were accompanied by alteration in depression‐related fecal microbiota and enhanced gut 5‐ HT metabolism, as well as elevated serum 5‐HT concentrations. We demonstrated that Maca‐EVs enhance the 5-HT-regulated BDNF expressions by activating the BDNF/TrkB/AKT axis. These data suggest that the regulation of the gut–brain axis by Maca‐EVs may be one element of its therapeutic mechanisms. Given the ability of PDEVs to cross the BBB, Maca‐EVs represent a promising platform for the management of depression. However, further studies are required to fully evaluate the safety and efficacy of this approach before clinical application.

AUTHOR CONTRIBUTIONS

Su‐Hua Qi and Bing Gu conceived and designed this study. Rui Hong, Lan Luo, Liang Wang, and Zhao‐Li Hu performed animal experiments and/or analyzed data and/or prepared the figures. Rong‐Qi Yin, Ming Li, Bin Gu, and Bin Wang analyzed data. Tao Zhuang and Xin‐ Yue Zhang performed animal experiments. Wan Wang, Yuan Zhou, and Lin‐Yan Huang assisted the study. Rui Hong, Lan Luo, and Liang Wang wrote the first version of the manuscript with input from coauthors. Su‐Hua Qi and Bing Gu revised the manuscript. All authors have read and approved the final manuscript.

ACKNOWLEDGMENTS

This work was supported by the Natural Science Foundation of Jiangsu Province (grant no. BK20211348), the Scientific Research Project of Jiangsu Provincial Healthy Commission (grant no. ZDB2020024), Jiangsu Qing‐Lan Project (Year 2020), the Leadership program (grant no. JBGS202203), and the Young Science and Technology Innovation Team of Xuzhou Medical University (grant no. TD202005 and TD202001).

ANTIDEPRESSANT EFFECTS OF MACA‐EVS \blacksquare

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

A data availability statement confirms the presence or absence of shared data. All data generated or analyzed during this study are included in this article and its supplementary files. Supplementary materials (figures, tables, scripts, graphical abstract, slides, videos, Chinese translated version, and update materials) may be found in the online DOI or iMeta Science [http://www.imeta.](http://www.imeta.science/) [science/](http://www.imeta.science/).

ETHICS STATEMENT

The ethics application (No. 202006W037) was approved by the Research Ethics Committee of Xuzhou Medical University.

ORCID

Bing Gu <http://orcid.org/0000-0001-7764-3213> Su‐Hua Qi <http://orcid.org/0000-0002-1479-701X>

REFERENCES

- 1. Santomauro, Damian F., Ana M. Mantilla Herrera, Jamileh Shadid, Peng Zheng, Charlie Ashbaugh, David M. Pigott, Cristiana Abbafati, et al. 2021. "Global Prevalence and Burden of Depressive and Anxiety Disorders in 204 Countries and Territories in 2020 Due to the COVID‐19 Pandemic." The Lancet 398: 1700–12. [https://doi.org/10.1016/](https://doi.org/10.1016/S0140-6736(21)02143-7) [S0140-6736\(21\)02143-7](https://doi.org/10.1016/S0140-6736(21)02143-7)
- 2. Thompson, Scott M., Angy J. Kallarackal, Mark D. Kvarta, Adam M. Van Dyke, Tara A. LeGates, and Xiang Cai. 2015. "An Excitatory Synapse Hypothesis of Depression." Trends in Neurosciences 38: 279–94. [https://doi.org/10.1016/j.tins.2015.](https://doi.org/10.1016/j.tins.2015.03.003) [03.003](https://doi.org/10.1016/j.tins.2015.03.003)
- 3. Yirmiya, Raz, Neta Rimmerman, and Ronen Reshef. 2015. "Depression as a Microglial Disease." Trends in Neurosciences 38: 637–58. <https://doi.org/10.1016/j.tins.2015.08.001>
- 4. Hodes, Georgia E., Veronika Kana, Caroline Menard, Miriam Merad, and Scott J. Russo. 2015. "Neuroimmune Mechanisms of Depression." Nature Neuroscience 18: 1386–93. <https://doi.org/10.1038/nn.4113>
- 5. Yohn, Christine N., Mark M. Gergues, and Benjamin Adam Samuels. 2017. "The Role of 5‐HT Receptors in Depression." Molecular Brain 10: 28. [https://doi.org/10.1186/](https://doi.org/10.1186/s13041-017-0306-y) [s13041-017-0306-y](https://doi.org/10.1186/s13041-017-0306-y)
- 6. David, D. J., and D. Gourion. 2016. "Antidépresseurs et tolérance: déterminants et prise en charge des principaux effets indésirables." L'Encéphale 42: 553–61. [https://doi.org/](https://doi.org/10.1016/j.encep.2016.05.006) [10.1016/j.encep.2016.05.006](https://doi.org/10.1016/j.encep.2016.05.006)
- 7. Bschor, Tom, Marcus Ising, Sebastian Erbe, Patricia Winkelmann, Dirk Ritter, Manfred Uhr, and Ute Lewitzka. 2012. "Impact of Citalopram on the HPA System. A Study of the Combined DEX/CRH Test in 30 Unipolar Depressed Patients." Journal of Psychiatric Research 46: 111–17. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jpsychires.2011.09.020) [jpsychires.2011.09.020](https://doi.org/10.1016/j.jpsychires.2011.09.020)

$\frac{16 \text{ of } 18 \text{ HONG ET AL.}}{160 \text{ N}}$

- 8. Chang, Lijia, Yan Wei, and Kenji Hashimoto. 2022. "Brain–Gut–Microbiota Axis in Depression: A Historical Overview and Future Directions." Brain Research Bulletin 182: 44–56. <https://doi.org/10.1016/j.brainresbull.2022.02.004>
- 9. Zhang, Jiancheng, Li Ma, Lijia Chang, Yaoyu Pu, Youge Qu, and Kenji Hashimoto. 2020. "A Key Role of the Subdiaphragmatic Vagus Nerve in the Depression‐Like Phenotype and Abnormal Composition of Gut Microbiota in Mice After Lipopolysaccharide Administration." Translational Psychiatry 10: 186. <https://doi.org/10.1038/s41398-020-00878-3>
- 10. Wang, Siming, Tamaki Ishima, Jiancheng Zhang, Youge Qu, Lijia Chang, Yaoyu Pu, Yuko Fujita, Yunfei Tan, Xingming Wang, and Kenji Hashimoto. 2020. "Ingestion of Lactobacillus intestinalis and Lactobacillus reuteri Causes Depression‐ and Anhedonia‐like Phenotypes in Antibiotic‐Treated Mice Via the Vagus Nerve." Journal of Neuroinflammation 17: 241. [https://doi.](https://doi.org/10.1186/s12974-020-01916-z) [org/10.1186/s12974-020-01916-z](https://doi.org/10.1186/s12974-020-01916-z)
- 11. Zheng, Danping, Timur Liwinski, and Eran Elinav. 2020. "Interaction Between Microbiota and Immunity in Health and Disease." Cell Research 30: 492–506. [https://doi.org/10.1038/](https://doi.org/10.1038/s41422-020-0332-7) [s41422-020-0332-7](https://doi.org/10.1038/s41422-020-0332-7)
- 12. Parker, Aimée, Sonia Fonseca, and Simon R. Carding. 2020. "Gut Microbes and Metabolites as Modulators of Blood–Brain Barrier Integrity and Brain Health." Gut Microbes 11: 135–57. <https://doi.org/10.1080/19490976.2019.1638722>
- 13. da Silva Leitão Peres, Natália, Letícia Cabrera Parra Bortoluzzi, Leila Larisa Medeiros Marques, Maysa Formigoni, Renata Hernandez Barros Fuchs, Adriana Aparecida Droval, and Flávia Aparecida Reitz Cardoso. 2020. "Medicinal Effects of Peruvian Maca (Lepidium meyenii): A Review." Food & Function 11: 83–92. <https://doi.org/10.1039/C9FO02732G>
- 14. Wang, Sunan, and Fan Zhu. 2019. "Chemical Composition and Health Effects of Maca (Lepidium meyenii)." Food Chemistry 288: 422–43. [https://doi.org/10.1016/j.foodchem.](https://doi.org/10.1016/j.foodchem.2019.02.071) [2019.02.071](https://doi.org/10.1016/j.foodchem.2019.02.071)
- 15. Yu, Zejun, Dong Li, Shengbing Zhai, Hang Xu, Hao Liu, Mingzhang Ao, Chunfang Zhao, Wenwen Jin, and Longjiang Yu. 2021. "Neuroprotective Effects of Macamide from Maca (Lepidium meyenii Walp.) on Corticosterone-Induced Hippocampal Impairments Through Its Anti‐ Inflammatory, Neurotrophic, and Synaptic Protection Properties." Food & Function 12: 9211–28. [https://doi.org/10.](https://doi.org/10.1039/D1FO01720A) [1039/D1FO01720A](https://doi.org/10.1039/D1FO01720A)
- 16. Rubio, Julio, Maria Caldas, Sonia Dávila, Manuel Gasco, and Gustavo F. Gonzales. 2006. "Effect of Three Different Cultivars of Lepidium meyenii (Maca) on Learning and Depression in Ovariectomized Mice." BMC Complementary and Alternative Medicine 6: 23. [https://doi.org/10.1186/1472-](https://doi.org/10.1186/1472-6882-6-23) [6882-6-23](https://doi.org/10.1186/1472-6882-6-23)
- 17. Ai, Zhong, Ai‐Fang Cheng, Yuan‐Tao Yu, Long‐Jiang Yu, and Wenwen Jin. 2014. "Antidepressant‐Like Behavioral, Anatomical, and Biochemical Effects of Petroleum Ether Extract from Maca (Lepidium meyenii) in Mice Exposed to Chronic Unpredictable Mild Stress." Journal of Medicinal Food 17: 535–42. <https://doi.org/10.1089/jmf.2013.2950>
- 18. Kameli, Nader, Anya Dragojlovic‐Kerkache, Paul Savelkoul, and Frank R. Stassen. 2021. "Plant‐Derived Extracellular Vesicles: Current Findings, Challenges, and Future Applications." Membranes 11: 411. <https://doi.org/10.3390/membranes11060411>
- 19. Ramos‐Zaldívar, Héctor M., Iva Polakovicova, Edison Salas‐ Huenuleo, Alejandro H. Corvalán, Marcelo J. Kogan, Claudia P. Yefi, and Marcelo E. Andia. 2022. "Extracellular Vesicles Through the Blood–Brain Barrier: A Review." Fluids and Barriers of the CNS 19: 60. [https://doi.org/10.1186/s12987-](https://doi.org/10.1186/s12987-022-00359-3) [022-00359-3](https://doi.org/10.1186/s12987-022-00359-3)
- 20. Verlinden, Heleen, Rut Vleugels, and Jozef Vanden Broeck. 2015. "Serotonin, Serotonin Receptors and Their Actions in Insects." Neurotransmitter 2: e314. [https://doi.org/10.14800/](https://doi.org/10.14800/nt.314) [nt.314](https://doi.org/10.14800/nt.314)
- 21. Mattson, Mark P., Stuart Maudsley, and Bronwen Martin. 2004. "BDNF and 5‐HT: A Dynamic Duo in Age‐Related Neuronal Plasticity and Neurodegenerative Disorders." Trends in Neurosciences 27: 589–94. [https://doi.org/10.1016/j.tins.2004.](https://doi.org/10.1016/j.tins.2004.08.001) [08.001](https://doi.org/10.1016/j.tins.2004.08.001)
- 22. Li, Chan, Junying Huang, Yung‐Chi Cheng, and Yuan‐Wei Zhang. 2020. "Traditional Chinese Medicine in Depression Treatment: From Molecules to Systems." Frontiers in Pharmacology 11: 586. [https://doi.org/10.3389/fphar.2020.](https://doi.org/10.3389/fphar.2020.00586) [00586;](https://doi.org/10.3389/fphar.2020.00586) [https://www.frontiersin.org/articles/10.3389/fphar.](https://www.frontiersin.org/articles/10.3389/fphar.2020.00586) [2020.00586](https://www.frontiersin.org/articles/10.3389/fphar.2020.00586)
- 23. Pino‐Figueroa, Alejandro, Diane Nguyen, and Timothy J. Maher. 2010. "Neuroprotective Effects of Lepidium meyenii (Maca): Lepidium meyenii Neuroprotective Effects." Annals of the New York Academy of Sciences 1199: 77–85. <https://doi.org/10.1111/j.1749-6632.2009.05174.x>
- 24. Wu, Hui, Charles J. Kelley, Alejandro Pino‐Figueroa, Huyen D. Vu, and Timothy J. Maher. 2013. "Macamides and Their Synthetic Analogs: Evaluation of In Vitro FAAH Inhibition." Bioorganic & Medicinal Chemistry 21: 5188–97. <https://doi.org/10.1016/j.bmc.2013.06.034>
- 25. Gonzales, Gustavo F., Sara Miranda, Jessica Nieto, Gilma Fernández, Sandra Yucra, Julio Rubio, Pedro Yi, and Manuel Gasco. 2005. "Red Maca (Lepidium meyenii) Reduced Prostate Size in Rats." Reproductive Biology and Endocrinology 3: 5. <https://doi.org/10.1186/1477-7827-3-5>
- 26. Lian, Meredith Qikai, Wei Heng Chng, Jeremy Liang, Hui Qing Yeo, Choon Keong Lee, Mona Belaid, Matteo Tollemeto, Matthias Gerhard Wacker, Bertrand Czarny, and Giorgia Pastorin. 2022. "Plant‐Derived Extracellular Vesicles: Recent Advancements and Current Challenges on Their Use for Biomedical Applications." Journal of Extracellular Vesicles 11: 12283. [https://doi.org/10.](https://doi.org/10.1002/jev2.12283) [1002/jev2.12283](https://doi.org/10.1002/jev2.12283)
- 27. Teng, Yun, Yi Ren, Mohammed Sayed, Xin Hu, Chao Lei, Anil Kumar, Elizabeth Hutchins, et al. 2018. "Plant‐Derived Exosomal MicroRNAs Shape the Gut Microbiota." Cell Host & Microbe 24: 637–652.e638. [https://doi.org/10.1016/j.chom.](https://doi.org/10.1016/j.chom.2018.10.001) [2018.10.001](https://doi.org/10.1016/j.chom.2018.10.001)
- 28. Nemati, Mohadeseh, Bipin Singh, Rakeeb Ahmad Mir, Mahdieh Nemati, Azadeh Babaei, Mahdi Ahmadi, Yousef Rasmi, Afsaneh Gholinejad Golezani, and Jafar Rezaie. 2022. "Plant‐Derived Extracellular Vesicles: A Novel Nanomedicine Approach with Advantages and Challenges." Cell Communication and Signaling 20: 69. [https://doi.](https://doi.org/10.1186/s12964-022-00889-1) [org/10.1186/s12964-022-00889-1](https://doi.org/10.1186/s12964-022-00889-1)
- 29. Cong, Minghui, Shenyu Tan, Simin Li, Lina Gao, Luqi Huang, Huang‐Ge Zhang, and Hongzhi Qiao. 2022. "Technology Insight: Plant‐Derived Vesicles—How Far From the Clinical

Biotherapeutics and Therapeutic Drug Carriers? Advanced Drug Delivery Reviews 182: 114108. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.addr.2021.114108) [addr.2021.114108](https://doi.org/10.1016/j.addr.2021.114108)

- 30. Cai, Heng, Lin‐Yan Huang, Rui Hong, Jin‐Xiu Song, Xin‐Jian Guo, Wei Zhou, and Zhao‐Li Hu, et al. 2022. "Momordica charantia Exosome‐Like Nanoparticles Exert Neuroprotective Effects Against Ischemic Brain Injury via Inhibiting Matrix Metalloproteinase 9 and Activating the AKT/GSK3β Signaling Pathway." Frontiers in Pharmacology 13: 908830. [https://doi.org/](https://doi.org/10.3389/fphar.2022.908830) [10.3389/fphar.2022.908830](https://doi.org/10.3389/fphar.2022.908830); [https://www.frontiersin.org/articles/](https://www.frontiersin.org/articles/10.3389/fphar.2022.908830) [10.3389/fphar.2022.908830](https://www.frontiersin.org/articles/10.3389/fphar.2022.908830)
- 31. Cui, Wen‐Wen, Cong Ye, Kai‐Xuan Wang, Xu Yang, Pei‐Yan Zhu, Kan Hu, Ting Lan, et al. 2022. "Momordica. charantia‐ Derived Extracellular Vesicles‐Like Nanovesicles Protect Cardiomyocytes Against Radiation Injury via Attenuating DNA Damage and Mitochondria Dysfunction." Frontiers in Cardiovascular Medicine 9: 864188. [https://doi.org/10.3389/](https://doi.org/10.3389/fcvm.2022.864188) [fcvm.2022.864188.](https://doi.org/10.3389/fcvm.2022.864188); [https://www.frontiersin.org/articles/10.](https://www.frontiersin.org/articles/10.3389/fcvm.2022.864188) [3389/fcvm.2022.864188](https://www.frontiersin.org/articles/10.3389/fcvm.2022.864188)
- 32. Morais, Livia H., Henry L. Schreiber, and Sarkis K. Mazmanian. 2021. "The Gut Microbiota–Brain Axis in Behaviour and Brain Disorders." Nature Reviews Microbiology 19: 241–55. [https://doi.](https://doi.org/10.1038/s41579-020-00460-0) [org/10.1038/s41579-020-00460-0](https://doi.org/10.1038/s41579-020-00460-0)
- 33. Winter, Gal, Robert A. Hart, Richard P. G. Charlesworth, and Christopher F. Sharpley. 2018. "Gut Microbiome and Depression: What We Know and What We Need to Know." Reviews in the Neurosciences 29: 629–43. [https://doi.org/10.1515/](https://doi.org/10.1515/revneuro-2017-0072) [revneuro-2017-0072](https://doi.org/10.1515/revneuro-2017-0072)
- 34. Kelly, John R., Yuliya Borre, Ciaran O'Brien, Elaine Patterson, Sahar El Aidy, Jennifer Deane, Paul J. Kennedy, et al. 2016. "Transferring the Blues: Depression‐Associated Gut Microbiota Induces Neurobehavioural Changes in the Rat." Journal of Psychiatric Research 82: 109–18. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jpsychires.2016.07.019) [jpsychires.2016.07.019](https://doi.org/10.1016/j.jpsychires.2016.07.019)
- 35. Valles‐Colomer, Mireia, Gwen Falony, Youssef Darzi, Ettje F. Tigchelaar, Jun Wang, Raul Y. Tito, Carmen Schiweck, et al. 2019. "The Neuroactive Potential of the Human Gut Microbiota in Quality of Life and Depression." Nature Microbiology 4: 623–32. <https://doi.org/10.1038/s41564-018-0337-x>
- 36. Barandouzi, Zahra Amirkhanzadeh, Angela R. Starkweather, Wendy A. Henderson, Adwoa Gyamfi, and Xiaomei S. Cong. 2020. "Altered Composition of Gut Microbiota in Depression: A Systematic Review." Frontiers in Psychiatry 11: 541. [https://doi.org/10.3389/fpsyt.2020.00541;](https://doi.org/10.3389/fpsyt.2020.00541) [https://www.](https://www.frontiersin.org/articles/10.3389/fpsyt.2020.00541) [frontiersin.org/articles/10.3389/fpsyt.2020.00541](https://www.frontiersin.org/articles/10.3389/fpsyt.2020.00541)
- 37. Bharwani, Aadil, M. Firoz Mian, Jane A. Foster, Michael G. Surette, John Bienenstock, and Paul Forsythe. 2016. "Structural & Functional Consequences of Chronic Psychosocial Stress on the Microbiome & Host." Psychoneuroendocrinology 63: 217–27. [https://doi.org/10.](https://doi.org/10.1016/j.psyneuen.2015.10.001) [1016/j.psyneuen.2015.10.001](https://doi.org/10.1016/j.psyneuen.2015.10.001)
- 38. Farshim, Pamela, Gemma Walton, Bhismadev Chakrabarti, Ian Givens, Doug Saddy, Ian Kitchen, Jonathan R. Swann, and Alexis Bailey. 2016. "Maternal Weaning Modulates Emotional Behavior and Regulates the Gut–Brain Axis." Scientific Reports 6: 21958. <https://doi.org/10.1038/srep21958>
- 39. Galley, Jeffrey D., Michael C. Nelson, Zhongtang Yu, Scot E. Dowd, Jens Walter, Purnima S. Kumar, Mark Lyte, and Michael T. Bailey. 2014. "Exposure to a Social Stressor

Disrupts the Community Structure of the Colonic Mucosa-Associated Microbiota." BMC Microbiology 14: 189. [https://](https://doi.org/10.1186/1471-2180-14-189) doi.org/10.1186/1471-2180-14-189

- 40. Yang, Jian, Peng Zheng, Yifan Li, Jing Wu, Xunmin Tan, Jingjing Zhou, Zuoli Sun, et al. 2020. "Landscapes of Bacterial and Metabolic Signatures and Their Interaction in Major Depressive Disorders." Science Advances 6: eaba8555. [https://](https://doi.org/10.1126/sciadv.aba8555) doi.org/10.1126/sciadv.aba8555
- 41. Moncrieff, Joanna, Ruth E. Cooper, Tom Stockmann, Simone Amendola, Michael P. Hengartner, and Mark A. Horowitz. 2022. "The Serotonin Theory of Depression: A Systematic Umbrella Review of the Evidence." Molecular Psychiatry. <https://doi.org/10.1038/s41380-022-01661-0>
- 42. Sukoff Rizzo, Stacey J., Claudine Pulicicchio, Jessica E. Malberg, Terrance H. Andree, Gary P. Stack, Zoë A. Hughes, Lee E. Schechter, and Sharon Rosenzweig‐ Lipson. 2009. "5‐HT1A Receptor Antagonism Reverses and Prevents Fluoxetine‐Induced Sexual Dysfunction in Rats." The International Journal of Neuropsychopharmacology 12: 1045–53. <https://doi.org/10.1017/S1461145709000406>
- 43. Beurel, Eléonore, Marisa Toups, and Charles B. Nemeroff. 2020. "The Bidirectional Relationship of Depression and Inflammation: Double Trouble." Neuron 107: 234–56. <https://doi.org/10.1016/j.neuron.2020.06.002>
- 44. Grosse, Laura, Livia A. Carvalho, Annemarie J. M. Wijkhuijs, Silja Bellingrath, Tillmann Ruland, Oliver Ambrée, Judith Alferink, Thomas Ehring, Hemmo A. Drexhage, and Volker Arolt. 2015. "Clinical Characteristics of Inflammation‐ Associated Depression: Monocyte Gene Expression Is Age‐ Related in Major Depressive Disorder." Brain, Behavior, and Immunity 44: 48–56. <https://doi.org/10.1016/j.bbi.2014.08.004>
- 45. Ng, Ada, Wilson W. Tam, Melvyn W. Zhang, Cyrus S. Ho, Syeda F. Husain, Roger S. McIntyre, and Roger C. Ho. 2018. "IL‐1β, IL‐6, TNF‐α and CRP in Elderly Patients with Depression or Alzheimer's Disease: Systematic Review and Meta‐Analysis." Scientific Reports 8: 12050. [https://doi.org/10.](https://doi.org/10.1038/s41598-018-30487-6) [1038/s41598-018-30487-6](https://doi.org/10.1038/s41598-018-30487-6)
- 46. Guseva, Daria, Alexander Wirth, and Evgeni Ponimaskin. 2014. "Cellular mechanisms of the 5‐HT7 receptor‐mediated signaling." Frontiers in Behavioral Neuroscience 8: 306. [https://doi.](https://doi.org/10.3389/fnbeh.2014.00306) [org/10.3389/fnbeh.2014.00306;](https://doi.org/10.3389/fnbeh.2014.00306) [https://www.frontiersin.org/](https://www.frontiersin.org/articles/10.3389/fnbeh.2014.00306) [articles/10.3389/fnbeh.2014.00306](https://www.frontiersin.org/articles/10.3389/fnbeh.2014.00306)
- 47. Udo, Hiroshi, Iksung Jin, Joung‐Hun Kim, Hsiu‐Ling Li, Trisha Youn, Robert D. Hawkins, Eric R. Kandel, and Craig H. Bailey. 2005. "Serotonin‐Induced Regulation of the Actin Network for Learning‐Related Synaptic Growth Requires Cdc42, N‐WASP, and PAK in Aplysia Sensory Neurons." Neuron 45: 887–901. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.neuron.2005.01.044) [neuron.2005.01.044](https://doi.org/10.1016/j.neuron.2005.01.044)
- 48. Elena, Kvachnina, Liu Guoquan, Dityatev Alexander, Renner Ute, Dumuis Aline, W. Richter Diethelm, Dityateva Galina, Schachner Melitta, A. Voyno‐Yasenetskaya Tatyana, and G. Ponimaskin Evgeni. 2005. "5‐HT7 Receptor Is Coupled to Gα Subunits of Heterotrimeric G12‐Protein to Regulate Gene Transcription and Neuronal Morphology." The Journal of Neuroscience 25: 7821. [https://doi.org/10.1523/](https://doi.org/10.1523/JNEUROSCI.1790-05.2005) [JNEUROSCI.1790-05.2005](https://doi.org/10.1523/JNEUROSCI.1790-05.2005)
- 49. Errico, Monica, Robert A. Crozier, Mark R. Plummer, and Daniel S. Cowen. 2001. "5‐HT7 Receptors Activate the

$\frac{18 \text{ of } 18 \text{ HONG ET AL.}}{18 \text{ of } 18 \text{ HONG ET AL.}}$

Mitogen Activated Protein Kinase Extracellular Signal Related Kinase in Cultured Rat Hippocampal Neurons." Neuroscience 102: 361–67. [https://doi.org/10.1016/S0306-4522\(00\)00460-7](https://doi.org/10.1016/S0306-4522(00)00460-7)

- 50. Speranza, Luisa, Angela Chambery, Marina Di Domenico, Marianna Crispino, Valeria Severino, Floriana Volpicelli, Marcello Leopoldo, Gian Carlo Bellenchi, Umberto di Porzio, and Carla Perrone‐Capano. 2013. "The Serotonin Receptor 7 Promotes Neurite Outgrowth Via ERK and Cdk5 Signaling Pathways." Neuropharmacology 67: 155–67. [https://](https://doi.org/10.1016/j.neuropharm.2012.10.026) doi.org/10.1016/j.neuropharm.2012.10.026
- 51. Yao, Wei, Qianqian Cao, Shilin Luo, Lujuan He, Chun Yang, Jiaxu Chen, Qi Qi, Kenji Hashimoto, and Ji‐chun Zhang. 2022. "Microglial ERK‐NRBP1‐CREB‐BDNF Signaling in Sustained Antidepressant Actions of (R)‐Ketamine." Molecular Psychiatry 27: 1618–29. <https://doi.org/10.1038/s41380-021-01377-7>
- 52. Yang, Chun, Qian Ren, Youge Qu, Ji‐Chun Zhang, Min Ma, Chao Dong, and Kenji Hashimoto. 2018. "Mechanistic Target of Rapamycin–Independent Antidepressant Effects of (R)‐ Ketamine in a Social Defeat Stress Model." Biological Psychiatry 83: 18–28. <https://doi.org/10.1016/j.biopsych.2017.05.016>
- 53. Adachi, Megumi, Anita E. Autry, Melissa Mahgoub, Kanzo Suzuki, and Lisa M. Monteggia. 2017. "TrkB Signaling in Dorsal Raphe Nucleus Is Essential for Antidepressant Efficacy and Normal Aggression Behavior." Neuropsychopharmacology 42: 886–94. [https://doi.org/10.](https://doi.org/10.1038/npp.2016.201) [1038/npp.2016.201](https://doi.org/10.1038/npp.2016.201)
- 54. He, Yacong, Shuang Chen, Bun Tsoi, Shuhua Qi, Bing Gu, Zhenxing Wang, Cheng Peng, and Jiangang Shen. 2021. "Alpinia oxyphylla Miq. and Its Active Compound P‐Coumaric Acid Promote Brain‐Derived Neurotrophic Factor Signaling for Inducing Hippocampal Neurogenesis and Improving Postcerebral Ischemic Spatial Cognitive Functions." Frontiers in

Cell and Developmental Biology 8: 577790. [https://doi.org/10.](https://doi.org/10.3389/fcell.2020.577790) [3389/fcell.2020.577790](https://doi.org/10.3389/fcell.2020.577790).; [https://www.frontiersin.org/articles/](https://www.frontiersin.org/articles/10.3389/fcell.2020.577790) [10.3389/fcell.2020.577790](https://www.frontiersin.org/articles/10.3389/fcell.2020.577790)

- 55. Chakraborty, Chiranjib, Ashish Ranjan Sharma, Garima Sharma, and Sang‐Soo Lee. 2021. "Therapeutic Advances of miRNAs: A Preclinical and Clinical Update." Journal of Advanced Research 28: 127–38. [https://doi.org/10.](https://doi.org/10.1016/j.jare.2020.08.012) [1016/j.jare.2020.08.012](https://doi.org/10.1016/j.jare.2020.08.012)
- 56. Murphy, Daniel E., Olivier G. de Jong, Maarten Brouwer, Matthew J. Wood, Grégory Lavieu, Raymond M. Schiffelers, Pieter Vader. 2019. "Extracellular Vesicle‐Based Therapeutics: Natural Versus Engineered Targeting and Trafficking." Experimental & Molecular Medicine 51: 1–12. [https://doi.org/](https://doi.org/10.1038/s12276-019-0223-5) [10.1038/s12276-019-0223-5](https://doi.org/10.1038/s12276-019-0223-5)

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Hong, Rui, Lan Luo, Liang Wang, Zhao‐Li Hu, Qi‐Rong Yin, Ming Li, Bin Gu, et al. 2023. "Lepidium meyenii Walp (Maca)‐derived extracellular vesicles ameliorate depression by promoting 5‐HT synthesis via the modulation of gut–brain axis." iMeta e116. <https://doi.org/10.1002/imt2.116>