Edith Cowan University Research Online

Research outputs 2014 to 2021

3-1-2019

# Features of the metabolic syndrome in late adolescence are associated with impaired testicular function at 20 years of age

R. J. Hart

D. A. Doherty

T. A. Mori

L. A. Adams

R-C. Huang

See next page for additional authors

Follow this and additional works at: https://ro.ecu.edu.au/ecuworkspost2013

Part of the Reproductive and Urinary Physiology Commons

10.1093/humrep/dey371

This is an Author's Accepted Manuscript of: Hart, R. J., Doherty, D. A., Mori, T. A., Adams, L. A., Huang, R. C., Minaee, N., ... & Olynyk, J. K. (2019). Features of the metabolic syndrome in late adolescence are associated with impaired testicular function at 20 years of age. *Human Reproduction, 34*(3), 389–402. Available here This Journal Article is posted at Research Online. https://ro.ecu.edu.au/ecuworkspost2013/5796

# Authors

R. J. Hart, D. A. Doherty, T. A. Mori, L. A. Adams, R-C. Huang, N. Minaee, D. J. Handelsman, R. McLachlan, R. J. Norman, J. E. Dickinson, J. K. Olynyk, and L. J. Beilin

1	Features of the metabolic syndrome in late adolescence are associated with impaired testicular
2	function at 20 years of age.
3	Short title Metabolic syndrome and testicular function
4	
5	R.J. Hart <sup>1,2,11</sup> , D.A. Doherty <sup>1,3</sup> , T.A. Mori <sup>4</sup> , L.A. Adams <sup>5</sup> , R-C Huang <sup>6</sup> , N Minaee <sup>3</sup> , D.J. Handelsman <sup>7</sup> , R
6	McLachlan <sup>8</sup> , R.J. Norman <sup>9</sup> , J. Dickinson <sup>1</sup> , J.K. Olynyk <sup>10</sup> , L.J. Beilin <sup>4</sup> .
7	<sup>1</sup> Division of Obstetrics and Gynaecology, University of Western Australia (UWA), Perth, WA, Australia
8	<sup>2</sup> Fertility Specialists of Western Australia, Claremont, WA
9	<sup>3</sup> Women and Infants Research Foundation, Perth WA.
10	<sup>4</sup> Medical School, UWA, Royal Perth Hospital, WA.
11	<sup>5</sup> Medical School, UWA, Sir Charles Gairdner Hospital, Nedlands WA.
12	<sup>6</sup> Telethon Kids Institute, UWA, Perth
13	<sup>7</sup> ANZAC Research Institute, University of Sydney, Sydney NSW
14	<sup>8</sup> Hudson Institute, Monash Medical Centre, Melbourne
15	<sup>9</sup> Robinson Institute, Research Centre for Reproductive Health, School of Medicine, University of
16	Adelaide & FertilitySA, Adelaide
17	<sup>10</sup> Department of Gastroenterology, Fiona Stanley and Fremantle Hospital and School of Medical and
18	Health Sciences, Edith Cowan University, WA.
19	
20	<sup>11</sup> Correspondence to
21	Roger Hart
22	Division of Obstetrics & Gynaecology
23	King Edward Memorial Hospital
24	374 Bagot Road
25	Subiaco
26	WA 6008
27	Tel +618 93401322
28	
29	E-mail <u>roger.hart@uwa.edu.au</u>
30	

3132 Abstract

33 Study question: Are early signs of metabolic disorder in late adolescence associated with features of 34 impaired testicular function many years before the majority are seeking parenthood?

**Summary answer:** Adolescents with features of metabolic disorder, as manifest by ultrasound evidence of non-alcoholic fatty liver (NAFL) at 17 years or systemic markers of inflammation, and/or insulin resistance measured by homeostasis model of insulin resistance (HOMA-IR) at 20 years of age, have reduced testicular volume, total sperm output, serum testosterone (T) and inhibin B (inhB) concentrations and a higher serum follicle stimulating hormone concentration (FSH), at 20 years of age, in comparison to their peers without metabolic disorder.

What is known already: Controversial evidence suggests a recent decline in sperm production potentially linked to environmental influences, but its cause remains unclear. The concomitant increases in obesity and diabetes suggests that lifestyle factors may contribute to this decline in testicular function. Although obesity has been associated with adverse testicular function in some studies, it remains unclear whether poor testicular function reflects, or causes, poor metabolic health. If metabolic disorder were present in adolescence, prior to the onset of obesity, this may suggest that metabolic disorder may lead to impaired testicular function

48 **Study design, size, duration:** The Western Australian Pregnancy Cohort (Raine) Study is a 49 longitudinal study of children born in 1989-1991 who have undergone detailed physical assessments 50 since birth (1454 male infants born). The purpose of this current sub-study was to perform a testicular 51 assessment at 20 years of age (913 were contactable).

At 17 years of age 490 underwent a hepatic ultrasound examination, serum cytokine assessment (n=520) and a metabolic assessment (n=544). A further metabolic assessment was performed at 20 years (n=608). Testicular assessment was performed on 20 year participants who consented to inclusion; 609 had reproductive hormones measured, 404 underwent a testicular ultrasound and 365 produced a semen sample.

57 **Participants/materials, setting, methods:** Testicular volume was estimated by ultrasonography, and 58 semen analysis performed by WHO methods. Serum was analysed to determine concentrations of 59 luteinising hormone (LH), FSH, inhB by immunoassays and T by liquid chromatography-mass 60 spectrometry (LC-MS).

At 17 years of age a liver ultrasound examination was performed to determine the presence of NAFL, and serum analysed for the following cytokines; interleukin-18 (IL18), soluble tumour necrosis factor receptor 1 & 2 (sTNFR1, sTNFR2) concentrations.

64 At 17 and 20 years of age fasting blood samples were analysed for serum liver enzymes, insulin, 65 glucose, triglycerides (TG), total cholesterol, high density lipoprotein (HDL) and low density lipoprotein 66 (LDL) cholesterol, high sensitivity (hs) CRP, and uric acid. HOMA was calculated and insulin 67 resistance (IR) was defined by a HOMA >4, anthropometric data was collected and dual energy X-ray 68 absorptiometry (DEXA) measurement was performed for lean and total fat mass. As at this young age 69 the prevalence of metabolic syndrome was expected to be low, a two-step cluster analysis was used 70 using waist circumference, TGs, insulin, and systolic blood pressure to derive a distinct high-risk 71 group with features consistent with the metabolic syndrome.

Main results and the role of chance: Men who at age 17 years were at elevated cardiometabolic risk had lower concentrations of T (medians: 4.0ng/ml vs 4.9ng/ml) and inhB (medians: 193.2pg/ml vs 221.9pg/ml) (p<0.001 for both) compared to those within the low risk metabolic cluster. Furthermore, men with ultrasound evidence of NAFLD detected at 17 years (n=45, 9.8%) had reduced total sperm output (medians: 68.0 million vs 126.00 million, p=0.044), T (4.0ng/ml vs 4.7ng/ml, p=0.005) and inhB (209.1pg/ml vs 218.4pg/ml, p=0.032) concentrations at 20 years compared to men without NAFLD.

78 Men with higher concentrations of sTNFR1, at 17 years of age, had a lower sperm output and seminal 79 volume, and serum concentrations of inhB, with an increase in LH and FSH at 20 years of age (all 80 p<0.05 after adjustment for age, body mass index [BMI], abstinence and a history of cryptorchidism 81 and varicocele, cigarette smoking, alcohol and drug use). Similarly, serum T was lower in men with a 82 higher fasting serum insulin, hsCRP, HOMA and total fat mass, and higher in men with higher fasting 83 HDL, iron at 20 years of age (all p<0.05). Multivariable regression analysis, adjusting for age and BMI 84 at 20 years, cryptorchidism and presence of a varicocele examined the associations between NAFLD 85 (at 17 years), and HOMA-IR >4 and metabolic cluster (at 20 years) with reproductive hormone 86 concentrations at age 20 years, demonstrated that men in the high-risk metabolic cluster at 20 years had a lower serum T and inhB (both p=0.012), and HOMA-IR >4 was associated with a lower serum T
(p=0.002), .

Limitations, reasons for caution: This study is limited by the sample size and multiple comparisons, and causality cannot be proven from an observational study. Due to a three year interval between some metabolic assessments and assessment of testicular function, we cannot exclude an introduction of a bias into the study, as some of the participants and their testicular function will not have been fully mature at the 17 year assessment.

94 Wider implications of the findings: Irrespective of a proven causation, our study findings are 95 important in that a significant minority of the men, prior to seeking parenthood, presented co-existent 96 features of metabolic disorder and signs of testicular impairment. Of particular note is that the 97 presence of NAFLD at 17 years of age, although only present in a minority of men, was associated 98 with an almost 50% reduction in sperm output at 20 years of age, and that the presence of IR at 20 99 years was associated with a 20% reduction in testicular volume.

Study funding/competing interest(s) This study was supported by Australian NHMRC Grant Numbers CE and received support from the Raine Medical Research Foundation, The Telethon Kids Institute, University of Western Australia, Women and Infants Research Foundation, Curtin University and Edith Cowan University.

## 104 Disclosures

105 Authors disclosures; DAD, JED, NM, LAA, R-CH, TAM, JKO, LJB have nothing to declare. RJH is 106 Medical Director of Fertility Specialists of Western Australia, has equity interests in Western IVF, and 107 has received grant support from MSD, Merck-Serono and Ferring Pharmaceuticals. RMcL has equity 108 interests in the Monash IVF Group. RJN has equity interests in FertilitySA, and has received grant 109 support from Merck Serono and Ferring Pharmaceuticals. DJH has received institutional grant funding 110 (but no personal income) for investigator-initiated testosterone pharmacology studies from Lawley and 111 Besins Healthcare and has provided expert testimony to anti-doping tribunals and for testosterone 112 litigation.

113

114 **Key words:** Raine, metabolic, testicular function, semen, reproductive hormones, testicular volume.

#### 117 Introduction

There is an on-going debate in reproductive medicine as to whether there has been a general decline in sperm production in recent times, potentially linked to environmental influences (1-3). The parallel increase in the rates of lifestyle related disorders, such as obesity and diabetes (4), raises the possibility that lifestyle factors may contribute to any potential decline in sperm production. In populations of men seeking fertility treatment, obesity has been associated with adverse testicular function; such as reduced testicular volume, seminal volume, sperm output, sperm motility, serum testosterone concentration and sperm DNA damage (5-9), although this has been challenged (10).

125

The metabolic syndrome is a cluster of adverse cardiovascular features including central obesity, atherogenic dyslipidemia, insulin resistance, a prothrombotic state, elevated blood pressure and increased circulating proinflammatory markers. Some evidence exists for an association between the metabolic syndrome and impaired testicular function in sub-fertile men (9, 11). However, causality is unclear whether these disorders have a common origin in early life (12), or whether impaired testicular function may induce or result from the metabolic disorder.

132

With the increase in the prevalence of features of the metabolic syndrome in adolescent populations (13), many will have ultrasound evidence of a fatty liver (14). Non-alcoholic fatty liver disease (NAFLD) is the most prevalent chronic liver disorder, affecting almost 1 in 5 adolescents (15), and is a recognized antecedent of progressive liver disease and cardiometabolic disorder (16). Hyperinsulinemia, or the presence of a fatty liver (17), is associated with a reduction in hepatic synthesis of sex hormone binding globulin (SHBG), increasing the metabolic clearance of testosterone.

140

141 The metabolic syndrome is associated with a low grade inflammatory state, with increased C-reactive 142 protein (CRP) and production of inflammatory cytokines, such as, Interleukin- 6 (IL-6), tumour 143 necrosis factor - $\alpha$  (TNF- $\alpha$ ), and their receptors 1 and 2 (TNFR1 and TNFR2), and the production of 144 oxygen free radicals, all of which may impair sperm and testicular function (18, 19). We therefore 145 proposed that impaired testicular function may reflect or cause poor metabolic health.

Our study was driven by the question; whether or not in a young adult population, representative of the Western Australian population (20), early signs of metabolic disorder are associated with a profile of impaired testicular function, many years before the majority of men seek paternity. Hence our aim was to relate antecedent and concurrent markers of adverse cardiometabolic health, in adolescence and early adulthood, to markers of testicular function within men at 20 years of age from the Western Australian Pregnancy Cohort (Raine) Study.

153

#### 154 Materials and Methods

155 The Raine study

156 The Raine Study (www.rainestudy.org.au) was designed to measure the relationships between early 157 life events and subsequent health and behaviour. The study recruited 2900 women around 18 weeks 158 of gestation in 1989-91 (20, 21). 2868 children (including 1454 boys) born to 2804 mothers were 159 retained to form the Raine Study cohort, and were studied every 2-3 years into early adulthood, 160 including detailed cardiometabolic assessment at 17 and 20 years of age, and 423 men underwent 161 testicular assessment by ultrasound and / or semen examinations (20, 22). Ethical approval was 162 obtained from the University of Western Australia Human Research Ethics Committee, and all 163 participants provided informed written consent for all aspects of the study.

164

#### 165 **Testicular function assessment**

166 Clinical and testicular function assessment at 20 years of age

167 All male cohort members were invited to attend follow-up, which involved questionnaires, collection of 168 anthropometric data (n=687), and collection of blood for analysis of serum testosterone, luteinizing 169 hormone (LH), FSH and inhB concentrations (n=609). A testicular ultrasound examination was 170 performed (n=404), and a semen sample (n=365) analysed at Fertility Specialists of Western 171 Australia, as previously reported (22). Semen samples were analysed as per WHO semen manual 172 guidelines (23) including sperm concentrations (million per ml), total sperm output (million per 173 ejaculate), motility (%A grade + %B grade) and morphology. The sperm chromatin structural assay 174 (SCSA) was performed as described (24) with slight modifications. The DNA fragmentation index 175 represents the percentage of sperm within the sample with fragmented or damaged DNA. Serum inhB 176 concentration was measured by Gen II ELISA (Beckman Coulter Inc. Brea, CA); LH and FSH were

measured by ELISA (IBL International, Hamburg, Germany), and testosterone was measured by
 liquid chromatography-tandem mass spectrometry (LC-MS/MS) as described (25) (for further details

179 refer to supplementary methods [sM]). Testicular ultrasonography was performed as described (22),

180 and the volume of each testis calculated (26). Varicocele was defined as present when the maximal

181 venous diameter was over 3mm, and increased with the Valsalva manoeuver (27).

182

## 183 Metabolic assessments

184 1. Metabolic assessment at 17 years of age

185 a. Hepatic ultrasound

186 The methods of hepatic ultrasound examinations conducted among 587 cohort members at age 17

187 years for diagnosing NAFL have been reported previously, and the data was used in this study (sM)]

188 (14).

#### 189 b. Cytokine assessment

The serum from 520 cohort members was stored at -80 C and was analysed for the following cytokines; interleukin-18 (IL18) by ELISA, soluble tumor necrosis factor receptor 1 & 2 (sTNFR1, sTNFR2). Plasma IL-18 was quantitated with a commercially available ELISA method. Plasma sTNFR1 and sTNFR2 were quantified using cytometric Bead Array. Individual cytokine concentrations were determined using FCAP Array software (BD Biosciences) (sM).

195

# 196 c. Cardiometabolic assessment

197 Data from previous publication (13) was extracted for the fasting blood samples from 549 cohort 198 members which were analysed at the PathWest Laboratory at Royal Perth Hospital for serum liver 199 enzymes, insulin, glucose, triglycerides (TG), total cholesterol, HDL and LDL cholesterol, hsCRP, and 200 uric acid, as previously described (13, 29), excluding serum hsCRP concentrations >10mg/l (13, 29). 201 Glucose, insulin, total cholesterol and triglycerides were measured by automated analysers (sM). 202 HOMA was calculated by fasting insulin (microunits per milliliter) × fasting glucose (millimoles per 203 liter)]/22.5, and insulin resistance (IR) was defined by a HOMA >4 (30). Resting blood pressure (BP) 204 readings were taken (sM). The cardiometabolic data was used to derive a 'high risk metabolic cluster' 205 phenotyped previously in this cohort (13), and described below.

### 207 2. Cardiometabolic assessment at 20 years of age

Fasting blood samples from 620 cohort members at 20 years of age were analysed according to the same protocol for the 17 year cardiometabolic assessment. To assess body fat distribution DEXA measurement was performed (31, 32).

- 211
- 212

## 213 Statistical Considerations

#### 214 Derivation of metabolic cluster at 20 years of age

215 The two most frequently used definitions of the metabolic syndrome in adulthood are the National 216 Cholesterol Education Program expert panel on detection, evaluation, and treatment of high blood 217 cholesterol in adults (NCEP ATP-III) (33), and the International Diabetes Federation definition (IDF) 218 (34), which differ significantly on the components of their definition (see details in sM). Hence, as 219 there is no universally accepted definition, and it was expected that at this young age the prevalence 220 of metabolic syndrome would be low, an alternative approach, a two-step cluster analysis was used 221 (13, 35, 36). This is an effective tool used to define groups accounting for variables for where there is 222 strong evidence of clustering. Within a single cluster, the subjects are relatively homogeneous, 223 sharing similar traits and being dissimilar to subjects in other clusters. The technique uses a scalable 224 cluster analysis algorithm (37), designed specifically to handle large data sets and has been used 225 previously within this cohort (13, 35, 36). It preselects subjects into sub-clusters before further 226 grouping into the desired number of clusters with use of log-likelihood distance. The cluster groups 227 were formed with use of; waist circumference, TGs, insulin, and systolic BP measured at 20 years of 228 age to derive distinct high-risk group with features consistent with the metabolic syndrome. This 229 approach was used previously to identify those Raine study participants within a high risk metabolic 230 cluster at 17 years of age (13).

231

#### 232 Data analysis

233 Continuous data were summarized using medians and inter-quartile ranges (IQR), reported as Q1– 234 Q3, when following a non-Gaussian distribution. Categorical data were summarized using frequency 235 distributions. Multivariable linear regression analysis was used to examine associations between 236 metabolic parameters and reproductive outcomes or hormone concentrations. Covariate adjustments summarized using standardised coefficients ( $\beta$ ) and their 95% confidence intervals (CI). Effects of the metabolic parameters on outcomes were presented without ( $\beta_1$ ) and with ( $\beta_2$ ) adjustment for BMI. Supplementary analyses adjusting for waist circumference instead of BMI were performed with analogous results (data not shown). Reproductive outcomes had a non-Gaussian distribution, and were transformed to normality either via logarithmic or power transformations determined using the Box-Cox analysis.

included abstinence, history of cryptorchidism, varicoceles and BMI. Regression results were

244

237

Differences in reproductive parameters and hormone concentrations across low and high risk metabolic clusters, HOMA-IR, NAFLD, insulin and hsCRP were investigated univariately using Mann-Whitney test for two groups. When appropriate univariable analyses were supplemented with multivariable analyses to control for confounders age and BMI at 20 years, cryptorchidism and presence of a varicocele. All hypothesis tests were two-sided and *p*-values of <0.05 were considered statistically significant. No adjustments for multiple hypothesis testing were made in this exploratory study (38, 39). SPSS (version 22.0, IBM SPSS) statistical software was used for data analysis.

252

### 253 Results

#### 254 Demographics

255 Of 913 male cohort members who were contactable, 365 (40%) provided a semen sample and 256 represented 48% of the men who attended any of the assessments at 20 years of age and 404 257 underwent a testicular ultrasound and 609 had serum available for reproductive hormone assessment 258 (Table 1). Most (587) had undergone a liver ultrasound at 17 years of age and/or a fasting metabolic 259 assessment (544), and up to 608 had undergone some aspect of metabolic assessment at age 20 260 years of age (Table 2). Participants who took part in the testicular assessment (semen sample and/or 261 testicular ultrasound) were similar clinically to those that declined participation (Table 2). There was 262 no difference between the participants and the non-participants with respect to markers of socio-263 economic status (data not shown sM). The prevalence of metabolic syndrome among males within 264 the participants was 4.1% by the NCEP-ATPIII definition (40) and 5.4% using the IDF definition (34).

266 Associations between markers of metabolic disorder and testicular parameters

267

#### 268 Metabolic indices at 17 years.

269 Multivariable linear regression analysis adjusting for current age, abstinence, a history of 270 cryptorchidism, presence of a varicocele, and BMI revealed the association of semen parameters with 271 markers of systemic inflammation at 17 years of age: total sperm output was reduced in men at 20 272 years of age who had a higher serum IL18 (p=0.025), or sTNFR1 (p=0.036) and their sperm 273 concentration was negatively associated with their serum IL18 concentration (p=0.020) measured at 274 17 years (Table 3A). In addition higher sTNFR1 was negatively associated with inhibin B (p=0.011), 275 and positively associated with serum LH and FSH (p=0.015, and p=0.001 respectively) three years 276 later (Table 4). When adjustment was performed for waist circumference instead of BMI the results 277 were analogous (data not shown). We have previously shown that alcohol use, cigarette smoking and 278 recreational drug use in this cohort had no influence on markers of testicular function (22), and that 279 testicular volume was positively associated with height, and total soft and lean body mass(12)

280

Associations between metabolic cluster analysis at 17 years and subsequent testicular function at 20
 years of age

At 17 years of age 70 of 439 participants (15.9%) who would subsequently undergo the male reproductive assessment were clustered within the high metabolic risk group.

285

In an unadjusted analysis of the reproductive hormones of men within the high risk metabolic cluster at 17 years of age, had median T and inhB concentrations significantly lower (p<0.001 for both), in comparison to the men within the low risk metabolic cluster (Table 5).

289

### 290 Metabolic indices at 20 years.

After adjustment for age, abstinence, a history of cryptorchidism, varicocele and BMI; diastolic blood pressure and serum insulin at 20 years of age were negatively associated with testicular volume (p=0.028 and p=0.004 respectively), although diastolic blood pressure was positively associated with total sperm output (p=0.020) and seminal volume (p=0.014). ALT and GGT were positively associated with sperm morphology (p=0.008 and p=0.028 respectively) (Table 3A). 296

297 Associations between markers of metabolic disorder at 20 years of age and sex hormones 298 Multivariable regression analyses with adjustment for history of cryptorchidism and presence of a 299 varicocele shown that serum total testosterone (TT) was reduced in men with a higher fasting serum; 300 triglycerides, insulin, hsCRP, ferritin, ALT, HOMA score, and DEXA indices of fat when measured 301 concurrently at 20 years of age (Table 4) (all p<0.05). Serum TT was positively associated with serum 302 HDL cholesterol, iron and transferrin saturation (all p<0.05). After simultaneous adjustment for BMI 303 (and waist circumference-data not shown) positive associations with serum HDL, iron, transferrin 304 saturation and lean mass remained and a negative association with hsCRP and serum insulin 305 remained (all p<0.05).

306

307 Associations between metabolic cluster analysis at 20 years of age and testicular function

The number of men in the high risk metabolic cluster at 20 years of age, those insulin resistant as measured by the HOMA score and with NAFL, varied within the analyses due to the varying number of participants who took part in the various sub-studies (Table 1).

311

In an unadjusted analysis of the men within the high risk metabolic cluster at 20 years of age, their
median T and inhB concentrations were lower than men within the low risk metabolic cluster (Table
supplementary figures [sF] 1a and 1b).

315

## 316 Associations between HOMA as a proxy for IR at 20 years of age and testicular function

IR (as defined by a HOMA>4) was present in 24 out of 616 men (3.9%) at 20 years of age. In an unadjusted analysis, in comparison to those participants who were not IR, their median testicular volume was smaller, and median T and inhB concentrations were lower, and their median serum FSH concentration was higher (Table 6, sF2a-d). Furthermore the 51 men, out of 609 (8.4%), who had a fasting serum insulin greater than 10  $\mu$ U/ml (91<sup>st</sup> centile), at 20 years of age had lower median serum T and inhB concentrations, and their FSH concentration was greater (supplementary table [sT]1, sF3a-c).

324

325 Associations between presence of NAFLD at 17 years of age and subsequent testicular function

Ultrasound evidence of NAFLD was present in 44 men, out of 458 (9.6%) who subsequently underwent some assessment of testicular function. Compared to those participants without NAFLD, there were reductions in their; median total sperm output, serum T and inhB concentrations (sT2 and sF4a-c).

330

Associations between serum hsCRP either; above or below, the 75% percentile at 20 years of age
 and testicular function

333 Men whose serum hsCRP was greater than the 75% centile (1.62mg/l) at 20 years of age (after 334 exclusion of concentrations >10mg/l), in comparison to those below, had a reduction in their median 335 seminal volume, serum T, LH and FSH concentrations (sT3, sF5a-d).

336

337 *Multivariable analysis* 

338 Multivariable analysis demonstrated that being in the high risk metabolic cluster at 20 years of age 339 was associated with a lower serum testosterone and inhB, and HOMA-IR >4 was associated with a 340 lower serum testosterone concentration at 20 years (sT4).

341

# 342 Discussion

343 The findings of this observational study of adult men at 20 years of age, demonstrated that despite 344 the majority of men being of normal weight, a small minority already displayed features of metabolic 345 disturbance which are associated with adverse cardiovascular outcomes at a much older age. Men 346 with features of the metabolic syndrome, or who were IR at 20 years of age, or had ultrasound 347 evidence of NAFLD at 17 years of age, displayed a picture consistent with a degree of primary 348 hypogonadism as they had reductions in testicular volume, sperm output, and serum testosterone and 349 inhB, with a reciprocal increase in serum FSH at 20 years of age. All of these variables are well 350 established as adverse markers of reproductive potential (2, 22). Furthermore in considering potential 351 mechanisms for the observed finding it is possible there are contrasting influences of metabolic 352 disorder as; higher concentrations of the inflammatory markers sTNFR1 (and IL18 to a lesser 353 degree), when measured at 17 years were associated with subsequent reductions in sperm output, 354 seminal volume, sperm concentration, inhB, with reciprocal rises in LH and FSH, at 20 years of age, 355 suggesting a direct gonadotoxic effect of adolescent inflammation on subsequent testicular function,

and in contrast higher concentrations of hsCRP at 20 years of age had a potential central negative influence on serum FSH secretion (and LH to a lesser degree), inducing a central hypogonadal state with reductions in serum testosterone and seminal volume, however without a concomitant reduction inhB and testicular volume, these could be chance associations.

360

361 One can speculate that if their cardiometabolic picture deteriorates over time, their testicular function 362 might further deteriorate, leading to an adverse effect on their reproductive potential. Conversely if 363 their metabolic picture improves, this may have a positive impact on their reproductive potential, 364 although. it is interesting to note that already at 20 years of age, irrespective of BMI, the markers of 365 cardiometabolic disorder; a higher fasting serum insulin, triglycerides, hsCRP, HOMA score and 366 DEXA fat mass were negatively associated with the testicular hormones; serum testosterone and 367 inhB, concentrations, while a higher total lean mass, serum HDL cholesterol and iron stores were 368 positively associated with these hormones. These findings offer a potential link between metabolic 369 and reproductive health, in that these adverse metabolic features recorded at 17 and 20 years of age 370 may predispose a man to later impaired testicular function, irrespective of adiposity. Although the 371 direction of causality will require further investigation, as it is established that a low circulating 372 testosterone is associated with cardiometabolic disorder (41-43). However, data from one prospective 373 study suggests that a low serum testosterone may be a risk marker for the development of 374 cardiometabolic disorder, rather than a causative risk factor (44). Due to a three year interval between 375 some metabolic assessments and assessment of testicular function, we cannot exclude an 376 introduction of a bias into the study, as some of the participants and their testicular function will not 377 have been fully mature at the 17 year assessment, and it is known that pubertal maturation can have 378 a moderating impact on obesity associated inflammation (45). Irrespective of a proven causation, our 379 study findings are important in that a significant minority of the men, prior to seeking parenthood, 380 presented with some features of metabolic disorder and signs of testicular impairment.

381

These study findings warrant further study in other cohorts. Of particular note is that the presence of NAFLD at aged 17 years of age, although only present in a minority of men, was associated with an almost 50% reduction in sperm output at 20 years of age, and that the presence of IR at 20 years was associated with; a 20% reduction in testicular volume, a 30% reduction in serum testosterone, and a
20% reduction in serum inhB concentrations.

387

# 388 Conclusion

This study has demonstrated an association of adverse cardiometabolic features with impaired testicular function at 20 years of age. Furthermore, it is notable that, despite the majority of the young men having apparently normal metabolic function, a significant minority were already showing some features of the metabolic syndrome.

393

# 394 Acknowledgements

395 We are extremely grateful to The Raine Study participants and their families who took part in this 396 study and the Raine Study Team for cohort co-ordination and data collection, and the NH&MRC for 397 their long term contribution to funding the study over the last 28 years. We acknowledge the Raine 398 Medical Research Foundation, The Telethon Kids Institute, The University of Western Australia, Edith 399 Cowan University, Murdoch University, The University of Notre Dame Australia and Curtin University 400 for financial support and for providing funding for core management of the Raine Study. We are 401 particularly grateful to Alex D'Vauz and Jenny Mountain of the Raine Study Team, Michele Pedretti for 402 performing the ultrasound examinations and the staff of FSWA for facilitating the semen 403 assessments, and Professor David Handelsman's laboratory, the ANZAC institute, for the steroid 404 assays and for his academic input, and to the Canadian Institutes for Health Research (MOP82893) 405 and Stephen Lye for funding the DEXA scanning.

406

# 409 Author contributions (supplementary data on line)

# 412 References

Levine H, Jørgensen H, Martino-Andrade a, Mendiola J, Weksler-Derri D, Mindlis I, et al.
 Temporal trends in sperm count: a systematic review and meta-regression analysis, Human
 Reproduction Update. 2017.

Skakkebaek NE, Rajpert-De Meyts E, Buck Louis GM, Toppari J, Andersson AM, Eisenberg ML,
et al. Male Reproductive Disorders and Fertility Trends: Influences of Environment and Genetic
Susceptibility. Physiol Rev. 2016;96(1):55-97.

4193.Handelsman DJ, Cooper TG. Falling sperm counts and global estrogenic pollution: what have420we learned over 20 years? Asian J Androl. 2013;15(2):159-61.

421 4. Finucane MM, Stevens GA, Cowan MJ, Danaei G, Lin JK, Paciorek CJ, et al. National, regional,
422 and global trends in body-mass index since 1980: systematic analysis of health examination surveys
423 and epidemiological studies with 960 country-years and 9.1 million participants. Lancet.
424 2011;377(9765):557-67.

5. Belloc S, Cohen-Bacrie M, Amar E, Izard V, Benkhalifa M, Dalleac A, et al. High body mass index has a deleterious effect on semen parameters except morphology: results from a large cohort study. Fertil Steril. 2014;102(5):1268-73.

428 6. Ramlau-Hansen CH, Thulstrup AM, Nohr EA, Bonde JP, Sorensen TI, Olsen J. Subfecundity in 429 overweight and obese couples. Hum Reprod. 2007;22(6):1634-7.

4307.Hammoud AO, Gibson M, Peterson CM, Meikle AW, Carrell DT. Impact of male obesity on431infertility: a critical review of the current literature. Fertil Steril. 2008;90(4):897-904.

432 8. Kort HI, Massey JB, Elsner CW, Mitchell-Leef D, Shapiro DB, Witt MA, et al. Impact of body 433 mass index values on sperm quantity and quality. J Androl. 2006;27(3):450-2.

434 9. Ventimiglia E, Capogrosso P, Colicchia M, Boeri L, Serino A, Castagna G, et al. Metabolic 435 syndrome in white European men presenting for primary couple's infertility: investigation of the 436 clinical and reproductive burden. Andrology. 2016;4(5):944-51.

437 10. MacDonald AA, Herbison GP, Showell M, Farquhar CM. The impact of body mass index on
438 semen parameters and reproductive hormones in human males: a systematic review with meta439 analysis. Hum Reprod Update. 2010;16(3):293-311.

440 11. Eisenberg ML, Li S, Behr B, Pera RR, Cullen MR. Relationship between semen production and441 medical comorbidity. Fertil Steril. 2015;103(1):66-71.

Hart RJ, Doherty DA, Keelan JA, McLachlan R, Skakkebaek NE, Norman RJ, et al. Early Life
Events Predict Adult Testicular Function; Data Derived From the Western Australian (Raine) Birth
Cohort. J Clin Endocrinol Metab. 2016;101(9):3333-44.

Huang RC, Mori TA, Burke V, Newnham J, Stanley FJ, Landau LI, et al. Synergy between
adiposity, insulin resistance, metabolic risk factors, and inflammation in adolescents. Diabetes Care.
2009;32(4):695-701.

448 14. Ayonrinde OT, Olynyk JK, Beilin LJ, Mori TA, Pennell CE, de Klerk N, et al. Gender-specific
449 differences in adipose distribution and adipocytokines influence adolescent nonalcoholic fatty liver
450 disease. Hepatology. 2011;53(3):800-9.

45115.Schwimmer JB, Deutsch R, Kahen T, Lavine JE, Stanley C, Behling C. Prevalence of fatty liver452in children and adolescents. Pediatrics. 2006;118(4):1388-93.

453 16. Fazel Y, Koenig AB, Sayiner M, Goodman ZD, Younossi ZM. Epidemiology and natural history
454 of non-alcoholic fatty liver disease. Metabolism. 2016;65(8):1017-25.

Simo R, Saez-Lopez C, Barbosa-Desongles A, Hernandez C, Selva DM. Novel insights in SHBG
 regulation and clinical implications. Trends Endocrinol Metab. 2015;26(7):376-83.

457 18. Aitken RJ. Reactive oxygen species as mediators of sperm capacitation and pathological458 damage. Mol Reprod Dev. 2017.

459 19. Bobjer J, Katrinaki M, Tsatsanis C, Lundberg Giwercman Y, Giwercman A. Negative
460 association between testosterone concentration and inflammatory markers in young men: a nested
461 cross-sectional study. PLoS One. 2013;8(4):e61466.

462 20. Straker L, Mountain J, Jacques A, White S, Smith A, Landau L, et al. Cohort Profile: The 463 Western Australian Pregnancy Cohort (Raine) Study-Generation 2. Int J Epidemiol. 2017.

464 21. Newnham JP, Evans SF, Michael CA, Stanley FJ, Landau LI. Effects of frequent ultrasound
465 during pregnancy: a randomised controlled trial. Lancet. 1993;342(8876):887-91.

466 22. Hart RJ, Doherty DA, McLachlan RI, Walls ML, Keelan JA, Dickinson JE, et al. Testicular 467 function in a birth cohort of young men. Hum Reprod. 2015;30(12):2713-24.

23. Cooper TG, Noonan E, von Eckardstein S, Auger J, Baker HW, Behre HM, et al. World Health
Organization reference values for human semen characteristics. Hum Reprod Update.
2010;16(3):231-45.

471 24. Evenson D, Jost L. Sperm chromatin structure assay is useful for fertility assessment.
472 Methods Cell Sci. 2000;22(2-3):169-89.

473 25. Harwood DT, Handelsman DJ. Development and validation of a sensitive liquid 474 chromatography-tandem mass spectrometry assay to simultaneously measure androgens and 475 estrogens in serum without derivatization. Clin Chim Acta. 2009;409(1-2):78-84.

476 26. Sakamoto H, Saito K, Oohta M, Inoue K, Ogawa Y, Yoshida H. Testicular volume
477 measurement: comparison of ultrasonography, orchidometry, and water displacement. Urology.
478 2007;69(1):152-7.

479 27. Lenz S, Giwercman A, Elsborg A, Cohr KH, Jelnes JE, Carlsen E, et al. Ultrasonic testicular
480 texture and size in 444 men from the general population: correlation to semen quality. Eur Urol.
481 1993;24(2):231-8.

482 28. Hamaguchi M, Kojima T, Itoh Y, Harano Y, Fujii K, Nakajima T, et al. The severity of 483 ultrasonographic findings in nonalcoholic fatty liver disease reflects the metabolic syndrome and 484 visceral fat accumulation. The American journal of gastroenterology. 2007;102(12):2708-15.

Black LJ, Burrows S, Lucas RM, Marshall CE, Huang RC, Chan She Ping-Delfos W, et al. Serum
25-hydroxyvitamin D concentrations and cardiometabolic risk factors in adolescents and young
adults. Br J Nutr. 2016;115(11):1994-2002.

Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis
 model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin
 concentrations in man. Diabetologia. 1985;28(7):412-9.

491 31. Flegal KM, Shepherd JA, Looker AC, Graubard BI, Borrud LG, Ogden CL, et al. Comparisons of
492 percentage body fat, body mass index, waist circumference, and waist-stature ratio in adults. Am J
493 Clin Nutr. 2009;89(2):500-8.

494 32. Demmer DL, Beilin LJ, Hands B, Burrows S, Pennell CE, Lye SJ, et al. Dual Energy X-Ray
495 Absorptiometry Compared with Anthropometry in Relation to Cardio-Metabolic Risk Factors in a
496 Young Adult Population: Is the 'Gold Standard' Tarnished? PLoS One. 2016;11(9):e0162164.

497 33. Expert Panel on Detection E, And Treatment of High Blood Cholesterol In Adults. Executive
498 Summary of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on
499 Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III).
500 JAMA. 2001;285:2486-97.

50134.Alberti KG, Zimmet P, Shaw J. Metabolic syndrome--a new world-wide definition. A502Consensus Statement from the International Diabetes Federation. Diabet Med. 2006;23(5):469-80.

50335.Hart R, Doherty DA, Mori T, Huang RC, Norman RJ, Franks S, et al. Extent of metabolic risk in504adolescent girls with features of polycystic ovary syndrome. Fertil Steril. 2011;95(7):2347-53, 53 e1.

505 36. Huang RC, Mori TA, Burrows S, Le Ha C, Oddy WH, Herbison C, et al. Sex dimorphism in the 506 relation between early adiposity and cardiometabolic risk in adolescents. J Clin Endocrinol Metab. 507 2012;97(6):E1014-22.

508 37. Zhang T, Livny M, editors. BIRCH: An efficient data clustering method for very large 509 databases. Proceedings of the ACM SIGMOD Conference on Management of Data; 1996; Montreal: 510 ACM Press.

511 38. Wasserstein RL, Lazar NA. The ASA's statement on p-values: context, process, and purpose. 512 The American Statistician. 2016;70(2):129-33. 513 39. Lew MJ. Three inferential questions, two types of P-value. Supplementary material to the 514 ASA's statement on p-values. The American Statistician. 2016;70(2):129-33.

40. Expert Panel on Detection E, Treatment of High Blood Cholesterol in A. Executive Summary
of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on
Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III).
JAMA. 2001;285(19):2486-97.

519 41. Ding EL, Song Y, Malik VS, Liu S. Sex differences of endogenous sex hormones and risk of 520 type 2 diabetes: a systematic review and meta-analysis. JAMA. 2006;295(11):1288-99.

42. Araujo AB, Dixon JM, Suarez EA, Murad MH, Guey LT, Wittert GA. Clinical review:
Endogenous testosterone and mortality in men: a systematic review and meta-analysis. J Clin
Endocrinol Metab. 2011;96(10):3007-19.

43. Holmboe SA, Vradi E, Jensen TK, Linneberg A, Husemoen LL, Scheike T, et al. The Association of Reproductive Hormone Levels and All-Cause, Cancer, and Cardiovascular Disease Mortality in Men. J Clin Endocrinol Metab. 2015;100(12):4472-80.

44. Holmboe SA, Jensen TK, Linneberg A, Scheike T, Thuesen BH, Skakkebaek NE, et al. Low
Testosterone: A Risk Marker Rather Than a Risk Factor for Type 2 Diabetes. J Clin Endocrinol Metab.
2016;101(8):3180-90.

530 45. Mengel E, Tillmann V, Remmel L, Kool P, Purge P, Latt E, et al. Changes in inflammatory

531 markers in estonian pubertal boys with different BMI values and increments: A 3-Year Follow-Up

532 Study. Obesity (Silver Spring). 2017;25(3):600-7.

#### Supplementary Methods on line

#### Hepatic ultrasound examination

Ultrasonographers experienced in hepatic ultrasound performed the ultrasound examinations, as described (14), according to the protocol described by Hamaguchi (28), which provides 92% sensitivity and 100% specificity for the histological diagnosis of fatty liver (14). Boys with sonographic fatty liver and a self-reported weekly alcohol intake of less than 140g over the previous 12 months were classified as having NAFLD. Testing for hepatitis B or C virus infections was not performed due to low notification rates for these infections in local teenagers during the study period (14).

Sex hormone measurement; Serum inhibin B concentrations were measured in duplicate by Inhibin B Gen II ELISA from Beckman Coulter Inc. (Brea, CA), which had a limit of detection of 2.6 pg/ml. Luteinising hormone (LH), follicular stimulating hormone (FSH) levels were determined in duplicate using ELISA kits from IBL International, Hamburg, Germany. The limit of detection of the LH assay was 0.4 IU/L (calibrated against WHO IRP 80/552), while for FSH assay it was 0.2 IU/L (calibrated against NIBSC 92/510). The intra-assay precision (CV) of the ELISAs ranged from 8-11% based on the mean values for low and high value quality control samples from n=16-17 assays.

#### Measurement of serum inflammatory markers;

### Cytokine assessment

The serum from 520 cohort members was stored at -80 C and was analysed for the following cytokines; interleukin-18 (IL18) by ELISA (Medical Biological Laboratories, Nagoya, Japan), soluble tumor necrosis factor receptor 1 & 2 (sTNFR1, sTNFR2). Plasma IL-18 was quantitated with a commercially available ELISA method (Medical Biological Laboratories, Nagoya, Japan). Plasma sTNFR1 and sTNFR2 were quantified using cytometric Bead Array (CBA) Flex sets (BD PharMingen, San Diego, CA) on the BD FACSArrayTM bioanalyser (BD Biosciences, San Jose, California, USA). Procedures followed the manufacturer's recommendations. Individual cytokine concentrations were determined using FCAP Array software (BD Biosciences). The IL-18: Intra- and inter-assay coefficients of variation (CV) were 5.6% and 7.6%, respectively, with a sensitivity: 12.5 pg/MI.

c. Cardiometabolic assessment

Fasting blood samples from 454 cohort members were analysed at the PathWest Laboratory at Royal Perth Hospital for serum liver enzymes, insulin, glucose, triglycerides (TG), total cholesterol, high density lipoprotein (HDL) and low density lipoprotein (LDL) cholesterol, high sensitivity (hs) CRP, and uric acid, as previously described (14, 31), excluding serum hsCRP concentrations >10mg/l (14, 31). Glucose was measured by an automated Technicon Axon analyser (Bayer Diagnostics, Sydney, Australia) using a hexokinase method. Insulin was measured by automated radioimmunoassay (Tosoh, Tokyo, Japan). Total cholesterol and triglycerides were determined enzymatically on the Cobas MIRA analyzer (Roche Diagnostics) with reagents from Trace Scientific (Melbourne, Australia) (supplementary methods).

The intra-assay CV was 0.87% for total cholesterol, 1.92% for triglycerides, 1.04% for glucose, 1.78%. The intra- and inter-assay CV for insulin was 2.5%, sensitivity: 0.9 ng/MI. For leptin the intra and inter-assay CV were 3.3% and 5.4%, respectively, with a sensitivity of 7.8 pg/mL.

Resting blood pressure (BP) readings were taken using an oscillometric sphygmomanometer (DINAMAP vital signs monitor 8100, DINAMAP XL vital signs monitor, or DINAMAP ProCare 100), Soma Technology Bloomfiled, Connecticut) with the subject seated. The monitor was set to automatically record readings every 2 minutes and the mean of the second and third readings was calculated.

#### Sociodemographic assessment

Family income when children were aged 14 years was obtained by questionnaire from the primary caregiver, and the highest maternal education was obtained by questionnaire when children were aged 8 years (20).

# The commonest definitions of metabolic syndrome

The NCEP ATP III guidelines state that the metabolic syndrome may be diagnosed when a person has three or more of five components: central obesity (waist circumference), an elevated TG level, a reduced HDL-cholesterol level, an elevated BP or an elevated fasting glucose concentration (35). The

IDF definition requires a person to have central obesity plus any two of four additional factors: a raised TG level: a reduced HDL-cholesterol: a raised BP or a raised fasting plasma glucose (known type 2 diabetes) (36).

# Supplementary tables on-line

**Supplementary Table 1.** Comparison of testicular volume, semen parameters and serum hormones by serum insulin concentration above or below 10µu/ml, all assessments made at 20 years of age. Data are represented as Median (IQR, R).

Variables of interest	NHigh	<b>Insulin ≥ 10</b> μu/ml	Normal	Insulin < 10 μu/ml	p-value
Testicular volume (ml)	28	14.8 (12.5-16.6, 10.0-23.8)	345	15.1 (12.7-17.3, 7.6-28.4)	0.646
Semen parameters					
Volume (ml)	22	2.9 (1.6-4.1, 0.8-7.2)	317	2.8 (2.0-3.7, 0.1-11.0)	0.799
Total sperm output (M)	22	134.4 (44.3-239.0, 0-436)	317	110.5 (51.6-206.8, 0.0-	0.578
				927.5)	
Sperm concentration (M/ml)	22	45.5 (19.1-84.5, 0-160)	317	45.0 (22.5-70.5, 0-220)	0.795
SCSA (%)	21	2.9 (1.6-5.0, 1.1-10.8)	311	3.1 (1.8-5.2, 0.2-30.0)	0.658
Morphology (N, %)	21	6.0 (3.5-9.0, 2-17)	307	5 (3-7, 0-18)	0.159
Motility (a + b, %)	21	54.0 (44.5-69.0, 19-79)	314	58 (43-67, 1-88)	0.907
Serum hormones					
Testosterone (ng/mL)	51	3.6 (2.9-4.8, 1.1-7.1)	556	4.7 (3.7-5.9, 1.3-10.3)	<0.001
LH (IU/I)	51	9.9 (7.5-12.9, 5.6-18.8)	557	10.5 (8.3-13.1, 2.3-28.4)	0.287
FSH (IU/I)	51	5.2 (3.3-7.3, 1.1-25.8)	557	4.2 (2.9-6.0, 0.6-39.5)	0.018
InhB (pg/ml)	51	174.0 (136.7-222.3, 28.9-389.3)	558	218.6 (176.8-268.9, 4.5-	<0.001
		· · ·		543.9)	

**Supplementary Table 2.** Comparison of testicular volume, semen parameters and serum testicular hormones and gonadotrophins, assessed at 20 years of age, by ultrasound evidence of NAFLD, at 17 years of age. Data is represented as Median (IQR, R).

Outcome	NNAFLD	NAFLD	Normal	No NAFLD	p-value
Testicular volume (ml)	28	15.0 (12.6-17.2, 10.5-23.2)	270	14.8 (12.6-17.3, 7.6-28.4)	0.821
Sperm parameters					
Semen volume (ml)	25	2.5 (1.6-3.3, 0.7-6.2)	255	2.8 (1.9-3.8, 0.3-11.0)	0.161
Total sperm output (M)	25	68.0 (23.2-170.9, 0.0-551.8)	255	126.0 (55.8-213.0, 0.0-	0.044
				927.5)	
Sperm concentration (M/ml)	25	26 (15-69, 0-113)	255	47 (25-71, 0-220)	0.060
SCSA (%)	24	4.0 (1.8-5.9, 0.7-13.6)	249	3.0 (1.9-5.4, 0.2-30.0)	0.428
Sperm morphology (N, %)	24	5 (3-7, 1-17)	246	5 (3-7, 0-15)	0.979
Sperm motility (a + b, %)	24	56.5 (370-67.8, 6-82)	252	58 (43-67, 1-88)	0.591
Serum hormone concentrations					
Testosterone (ng/ml)	44	4.0 (3.2-5.0, 1.1-7.2)	391	4.7 (3.6-5.9, 1.3-9.9)	0.005
LH (IU/I)	44	8.8 (7.3-12.8, 5.4-18.9)	393	10.7 (8.6-13.4, 2.3-28.4)	0.054
FSH (IU/I)	44	3.6 (2.7-5.6, 1.2-14.3)	393	4.3 (3.1-6.3, 0.6-39.5)	0.428
InhB (pg/ml)	44	209.1 (145.85-253.2, 48.7-	393	218.4 (179.7-270.5, 4.5-	0.032
		389.3)		543.9)	

**Supplementary Table 3.** Comparison of testicular volume, semen parameters and serum hormones by serum hsCRP concentration by 75% percentile, all assessments made at 20 years of age. Data are represented as Median (IQR, R). Individuals with hsCRP>10 were excluded.

Variables of interest	$N_{High}$	hsCRP ≥ 1.615 mg/l	Normal	hsCRP < 1.615 mg/l	p-value
Testicular volume (ml)	60	15.4 (12.2-17.1, 8.9-23.5)	299	14.9 (12.9-17.2, 7.6-28.4)	0.955
Semen parameters					
Volume (ml)	53	2.5 (1.6-3.1, 0.7-6.8)	275	3.0 (2.0-3.8, 0.1-11.0)	0.043
Total sperm output (M)	53	84.8 (49.1-166.9, 0.0-639.2)	275	118.4 (50.4-214.6, 0.0-927.5)	0.127
Sperm concentration (M/ml)	53	35 (23-63, 0-180)	275	47.0 (21.5-74.0, 0-220)	0.209
SCSA (%)	51	2.9 (1.7-5.1, 0.7-10.8)	271	3.1 (1.9-5.4, 0.2-30.0)	0.606
Morphology (N, %)	51	5 (4-8, 2-13)	267	5 (3-7, 0-18)	0.239
Motility (a + b, %)	52	52.5 (40.3-65.8, 10.0-86.0)	273	58 (43-67, 1-88)	0.276
Serum hormones					
Testosterone (ng/ml)	126	4.0 (3.2-5.2, 1.1-8.1)	456	4.8 (3.8-6.0, 1.7-9.9)	<0.001
LH (IU/I)	126	9.5 (7.7-12.3, 4.3-19.5)	458	10.7 (8.6-13.2, 2.3-28.4)	0.008
FSH (IU/I)	126	3.9 (2.7-5.2, 0.6-14.3)	458	4.4 (3.0-6.4, 0.6-39.5)	0.024
InhB (pg/ml)	126	216.3 (156.0-260.2, 48.7-419.1)	458	216.2 (174.2-269.7, 4.5-543.9)	0.201

**Supplementary Table 4** Multivariate regression analysis to derive which metabolic risk factor individually remains significant, after adjustments for age and BMI at 20 years, cryptorchidism, and presence of a varicocele, on the reproductive hormones measured at 20 years of age. Three separate models, with either; HOMA at 20 years, NAFLD or metabolic cluster at 20 years were examined for each outcome. Model summaries include univariable and multivariable (adjusted) regression coefficients with their corresponding 95% confidence intervals (95% CI) and p-values; together with R<sup>2</sup> obtained in univariable models and an increase in R<sup>2</sup> (denoted R<sup>2</sup> change) in adjusted models are shown. P-value for R<sup>2</sup> change is reported to indicate whether the simultaneous adjustment for other characteristics improves prediction of outcomes. All measurements other than assessment for presence of NAFL (assessed at 17 years of age), were taken at 20 years of age.

Outcome (Total n=260)	Univariable			Multivariable					
· · · · ·	β (95% Cl)	<i>p</i> -value	R <sup>2</sup>	Adjusted β (95% CI)	<i>p</i> -value	R <sup>2</sup> change	<i>p</i> -value for		
Testosterone									
HOMA-IR	-0.54 (-0.85, -0.24)	<0.001	0.036	-0.48 (-0.78,-0.18)	0.002	0.063	<0.001		
NAFLD	-0.24 (-0.48, 0.009)	0.004	0.014	-0.12 (-0.37, 0.14)	0.364	0.050	0.010		
Metabolic cluster	-0.43 (-0.62, -0.25)	<0.001	0.060	-0.28 (-0.50, -0.06)	0.012	0.026	0.056		
FSH									
HOMA-IR	0.01 (-0.001, 0.034)	0.061	0.004	0.013 (-0.007, 0.034)	0.202	0.012	0.375		
NAFLD	0.002 (-0.01, 0.01)	0.667	0.000	0.02 (-0.02, 0.01)	0.842	0.013	0.487		
Metabolic cluster	0.003 (-0.01, 0.01)	0.547	0.002	0.01 (-0.01, 0.02)	0.546	0.015	0.275		
Inhibin B									
HOMA-IR	-10.5 (-22.9, -6.15)	0.097	0.008	-8.7 (-20.8, 3.44)	0.160	0.074	<0.001		
NAFLD	-3.0 (-12.7, 6.7)	0.545	0.001	1.1 (-8.9, 11.1)	0.829	0.075	0.001		
Metabolic cluster	-11.3 (-22.0, -2.5)	<0.001	0.042	-11.3 (-22.0, -2.5)	0.012	0.050	0.001		

NAFL = Non-alcoholic fatty liver diagnosed on ultrasound

HOMA = homeostasis model assessment insulin

**Table 1.** Flow of study participants. Total number of participants with measurements available are shown as (n=maximum number of participants) and the maximal number of participants for each outcome out of testicular volume, semen sample and blood sample assessment according to measurements taken during the various follow-ups listed. The presence of non-alcoholic fatty liver (NAFL) was derived from a previous study Ayonirynde et al (14) and the data identifying individuals within or without the high cardiometabolic risk cluster, derived using cluster analysis in a previous study Huang et al (13)

				N
Pregnant women enrolled in the Raine stud	dy			2900
Live births				2868
Male infants				1454
Female infants				1414
Male participants who had at least one of		Testicular volume	Semen sample	Serum available for
testicular ultrasound, semen or blood		assessment	provided	reproductive
samples (n=648)		performed	(n=365)	hormones
		(n=404)		(n=609)
Participants who underwent 16/17 ±	Total	Testicular	Semen sample	Serum sample
20/21 follow-up	participants	volume	parameters	available for
	underwent	measurements	available	gonadotrophins and
	assessment	available	(n)	testosterone
	(n)	(n)		(n)
16/17 year follow up				
Serum cytokines assessment	(n=520)	319	290	478
Liver ultrasound for NAFL	(n=587)	298	280	437
presence(14)				
Serum available for full metabolic	(n=544)	289	264	439
assessment (13)				
20/21 year follow up				
Contactable	(n=913)			
Participated	(n=705)			
Anthropometric examination	(n=687)			
Blood pressure measured	(n=693)	391	360	603
Serum available for biochemistry	(n=620)	374	340	609
Serum available for full metabolic	(n=608)	367	337	599
assessment				
Serum for HOMA calculation	(n=618)	373	339	609
DEXA scan performed	(n=634)	362	333	557

NAFL = Non-alcoholic fatty liver diagnosed on ultrasound

HOMA = homeostasis model assessment insulin (fasting insulin [µu/ml] × fasting glucose [mM]/22.5).

DEXA = Dual energy X-ray absorptiometry

Table 2. Partic	ipant characteristic	s at 20 years of ag	je – comparison l	between thos	e who participated	in at least one a	spect
of the testicula	r assessment and t	hose who did not.	Unless otherwise	e specified, da	ata were collected a	at 20 years of ag	le.

	Male n=64	participants 8	Male n=57	non-participants	<i>p</i> -value
	N	Median (IQR, R) or N (%)	N	Median (IQR, R) or N (%)	_
Age at 17 year follow-up	487	17.0 (16.9-17.1, 16.3-18.0)	33	17.0 (16.9-17.1, 16.7-17.3)	0.598
Age at 20 year follow-up	648	19.9 (19.7-20.3, 19.3-22.1)	57	19.9 (19.6-20.5, 19.4-21.7)	0.786
Anthropometric					
Height (cm)	632	180 (170-180, 162-199)	55	180 (180-190, 156-198)	0.550
Weight (kg)	632	75.9 (68.3-86.2, 52.2-137.5)	55	75.8 (69.0-86.1, 50.2-176.5)	0.884
BMI (kg/m²)	632	23.6 (21.4-26.3, 16.7-48.9)	55	23.9 (21.5-25.5, 18.0-42.9)	0.887
under 25		405 (64.1%)		39 (70.9%)	0.543
25 - 30		155 (24.5%)		10 (18.2%)	
30 plus		72 (11.4%)		6 (10.9%)	
Waist circumference (cm)	632	80.5 (75.1-87.5, 43.8-145.5)	55	80.8 (74.3-88.3, 63.5-131.5)	0.995
Adiposity (DEXA)					
l otal fat mass (g)	586	14935 (10519-2431, 3413- 105957)	48	15349 (10604-20633, 6583- 50244)	0.843
Total lean mass (g)	586	56702 (52052-61561, 33747-	48	57479 (51526-63403, 38679-	0.790
		83318)		89622)	
Soft tissue percentage <sup>a</sup>	586	21 (16-28, 6-63)	48	20 (16-28, 10-46)	0.771
Total fat percentage <sup>b</sup>	586	20 (15-27, 5-61)	48	19 (15-27, 10-45)	0.740
Biochemistry					
Fasting glucose (mmol/l)	616	5.0 (4.8-5.3, 3.1-8.2)	2	-	
Triglycerides (mmol/l)	616	1.0 (0.7-1.3, 0.3-17.8)	2	-	
HDL cholesterol (mmol/l)	616	1.2 (1.0-1.4, 0.6-2.6)	2	-	
LDL cholesterol (mmol/l)	616	2.4 (1.9-2.8, 0.2-5.3)	2	-	
Iron (umol/L)	617	16.1 (12.8-20.5, 3.0-40.7)	2	-	
I ransferrin (umol/l)	617	31.6 (29.0-34.0, 21.2-46.5)	2	-	
I ransferrin saturation (%)	617	26.2 (20.7-32.8, 4.7-84.1)	2	-	
Ferritin (ug/l)	392	87.8 (61.9-127.2, 6.3-326.9)	2	-	
Insuin (µu/mi)	616	2.0(2.0-4.7, 2.0-64.3)	2	-	
ALT ()	591	0.0(0.3-1.4, 0.1-9.8)	2	-	
	616	30(22-42, 10-372)	2	-	
	616	17(14-23, 7-03)	2	-	
Adipopectin (mg/l)	616	Z5 (ZZ-51, 11-199) 7 6 (5 1-10 3 0 6-34 6)	2	-	
Leptin (ug/l)	616	3.3(1.7.70, 0.162, 1)	2	-	
	616	0.5(0.4, 1, 1, 0, 2, 16, 2)	2	-	
	616	21 (3.9%)	2	-	
Metabolic clusters	010	24 (0.070)	2	0	
High risk at 20vrs	606	76 (12,5%)	2	0	
High risk at 17 yrs	439	70 (15.9%)	15	2 (13.3%)	
Blood pressure					
Systolic (mm/Hg)	636	122 (114-132, 90-160)	57	123 (112-131, 91-152)	0.792
Diastolic (mm/Hg)	636	65 (59-71, 46-96)	57	64 (60-69, 47-90)	0.609
Serum reproductive hormones					
Testosterone (ng/mL)	607	4.6 (3.6-5.8, 1.1-10.3)			
LH (iu/l)	608	10.5 (8.3-13.0, 2.3-28.4)			
FSH (iu/l)	608	4.3 (3.0-6.2, 0.6-39.5)			
InhB (pg/ml)	609	216.4 (170.4-266.4, 4.5-543.9)			
Cytokines(at 17 yrs)					
IL18 (pg/ml)	496	288.4 (231.2-373.9, 0-3122)	23	263.6 (236.4-363.1, 153-1109)	0.802
sINFR1 (pg/ml)	497	364.4 (286.1-462.8, 11-3549)	23	362.2 (293.7-420.4, 189-668)	0.874
sTNFR2 (pg/ml)	497	3180.4 (2636.3-3930.8, 24-	23	3222.3 (2588.0-4057.2, 1930-	0.853
Henatic ultrasound (at 17 ura)		9100)		5151)	
	150	45 (0.8%)	31	1 (12.0%)	0.757
	-53	+0 (0.070)	51	Ŧ (12.370)	0.101
Smoking <sup>A</sup>	494	78 (15.8%)	38	34 (15.8%)	1,000
Alcohol consumption <sup>^</sup>		- ( )			
Nil	492	85 (17.3%)	37	6 (16.2%)	0.923
Moderate		249 (50.6%)	-	11 (29.7%́)	
Binge		158 (̀32.1%)́		58 (28.2%)	

<sup>a</sup> Total soft tissue fat percentage = fat mass x 100 / (fat mass + lean mass), <sup>b</sup> Total fat percentage = fat mass x 100 / (fat mass + lean mass + bone mineral content), <sup>c</sup> hsCRP>10 has been excluded. <sup>A</sup>Smoking has 154 missing in the participants and 19 in the non-participants group. Alcohol consumption has 156 missing in the participants and 20 missing in the non-participants group.

Replaced with new Table 4

	Testis volume		Semen volume		Sperm output		Semen concentration	
	β₁ (95% CI)	β2 (95% CI)	β1 (95% CI)	β <sub>2</sub> (95% CI)	β1 (95% CI)	β2 (95% CI)	β1 (95% CI)	β2 (95% CI)
Biochemistry								
Glucose	-0.043 (-0.143, 0.057)	-0.057 (-0.157, 0.044)	0.066 (-0.034, 0.166)	0.078 (-0.023, 0.179)	0.071 (-0.026, 0.168)	0.085 (-0.013, 0.182)	0.057 (-0.045, 0.159)	0.065 (-0.038, 0.168)
Triglycerides	-0.047 (-0.147, 0.053)	-0.070 (-0.173, 0.032)	-0.044 (-0.144, 0.057)	-0.029 (-0.132, 0.074)	-0.044 (-0.141, 0.053)	-0.027 (-0.127, 0.004)	-0.012 (-0.114, 0.090)	-0.003 (-0.108, 0.102)
HDL cholesterol	-0.027 (-0.126, 0.073)	-0.002 (-0.106, 0.103)	0.030 (-0.070, 0.130)	0.010 (-0.095, 0.115)	0.027 (-0.070, 0.124)	0.003 (-0.098, 0.105)	-0.010 (-0.092, 0.111)	-0.003 (-0.110, 0.103)
LDL cholesterol	0.028 (-0.073, 0.128)	0.009 (-0.094, 0.112)	-0.023 (-0.123, 0.078)	-0.007 (-0.111, 0.096)	-0.001 (-0.099, 0.096)	0.018 (-0.082, 0.118)	0.015 (-0.087, 0.118)	0.026 (-0.079, 0.131)
Iron	-0.019 (-0.119, 0.081)	-0.017 (-0.117, 0.083)	0.092 (-0.008, 0.192)	0.090 (-0.010, 0.191)	-0.012 (-0.010, 0.086)	-0.014 (-0.111, 0.084)	-0.059 (-0.161, 0.043)	-0.060 (-0.162, 0.042)
Transferrin	0.081 (-0.019, 0.180)	0.072 (-0.028, 0.172)	0.020 (-0.080, 0.120)	0.029 (-0.072, 0.129)	-0.067 (-0.167, 0.033)	-0.057 (-0.153, 0.040)	-0.075 (-0.177, 0.026)	-0.071 (-0.174, 0.031)
Transferrin saturation %	-0.046 (-0.147, 0.054)	-0.041 (-0.141, 0.059)	0.076 (-0.025, 0.177)	0.072 (-0.029, 0.173)	-0.001 (-0.098, 0.097)	-0.006 (-0.103, 0.092)	-0.039 (-0.141, 0.064)	-0.042 (-0.145, 0.061)
Ferritin	-0.040 (-0.141, 0.060)	-0.057 (-0.159, 0.045)	-0.049 (-0.150, 0.053)	-0.037 (-0.140, 0.066)	-0.060 (-0.158, 0.037)	-0.047 (-0.147, 0.052)	-0.031 (-0.133, 0.072)	-0.024 (-0.128, 0.081)
Insulin	-0.116 (-0.215, -0.016)	-0.153 (-0.256, -0.049)	-0.036 (-0.137, 0.065)	-0.017 (-0.122, 0.088)	-0.013 (-0.111, 0.084)	0.011 (-0.091, 0.113)	0.001 (-0.101, 0.104)	0.015 (-0.092, 0.122)
hsCRP <sup>†</sup>	-0.003 (-0.105, 0.099)	-0.029 (-0.135, 0.077)	-0.113 (-0.216, -0.010)	-0.101 (-0.208, 0.007)	-0.065 (-0.164, 0.035)	-0.046 (-0.149, 0.057)	-0.046 (-0.150, 0.058)	-0.037 (-0.146, 0.071)
ALT	-0.034 (-0.135, 0.066)	-0.061 (-0.165, 0.043)	-0.106 (-0.207, -0.006)	-0.093 (-0.202, 0.016)	-0.008 (-0.105, 0.090)	0.015 (-0.086, 0.117)	0.024 (-0.078, 0.127)	0.039 (-0.068, 0.145)
AST	-0.010 (-0.112, 0.092)	-0.041 (-0.149, 0.066)	-0.031 (-0.133, 0.072)	0.009 (-0.117, 0.098)	-0.045 (-0.143, 0.054)	-0.021 (-0.125, 0.082)	-0.031 (-0.135, 0.073)	-0.019 (-0.128, 0.090)
GGT	0.003 (-0.097, 0.103)	-0.003 (-0.103, 0.097)	-0.021 (-0.121, 0.080)	-0.015 (-0.116, 0.085)	-0.017 (-0.115, 0.080)	-0.011 (-0.108, 0.086)	-0.018 (-0.119, 0.084)	-0.014 (-0.117, 0.088)
Blood pressure								
Systolic	0.005 (-0.097, 0.103)	-0.028 (-0.132, 0.076)	0.094 (-0.003, 0.191)	0.135 (0.032, 0.238)	0.006 (-0.089, 0.100)	0.038 (-0.062, 0.139)	-0.039 (-0.138, 0.060)	-0.028 (-0.134, 0.078)
Diastolic	-0.109 (-0.207, -0.011)	-0.124 (-0.223, -0.025)	0.112 (0.014, 0.209)	0.124 (0.026, 0.223)	0.100 (0.005, 0.194)	0.114 (0.018, 0.209)	0.049 (-0.050, 0.149)	0.057 (-0.044, 0.157)
Cytokines at 16/17 yrs								
IL18	0.022 (-0.088, 0.132)	0.026 (-0.084, 0.135)	0.003 (-0.107, 0.112)	-0.001 (-0.110, 0.109)	-0.116 (-0.222, -0.011)	-0.120 (-0.225, -0.015)	-0.129 (-0.239, -0.019)	-0.131 (-0.242, -0.011)
sTNFR1	-0.089 (-0.197, 0.020)	-0.094 (-0.203, 0.014)	-0.123 (-0.232, -0.015)	-0.118 (-0.228, -0.009)	-0.119 (-0.224, -0.013)	-0.113 (-0.219, -0.007)	-0.045 (-0.157, 0.066)	-0.043 (-0.155, 0.068)
sTNFR2	-0.009 (-0.118, 0.101)	-0.013 (-0.122, 0.096)	-0.084 (-0.193, 0.026)	-0.081 (-0.190, 0.029)	-0.079 (-0.185, 0.027)	-0.076 (-0.181, 0.030)	-0.065 (-0.177, 0.048)	-0.063 (-0.176, 0.049)
DEXA								
Total fat %	-0.018 (-0.122, 0.086)	-0.129 (-0.266, 0.007)	-0.086 (-0.189, 0.018)	-0.068 (-0.205, 0.070)	-0.096 (-0.199, 0.007)	-0.073 (-0.206, 0.059)	-0.045 (-0.150, 0.060)	-0.029 (-0.169, 0.111)
Soft tissue fat %	-0.035 (-0.139, 0.069)	-0.137 (-0.266, -0.008)	-0.084 (-0.187, 0.020)	-0.064 (-0.201, 0.073)	-0.095 (-0.195, 0.005)	-0.072 (-0.205, 0.060)	-0.046 (-0.151, 0.060)	-0.030 (-0.170, 0.109)
Total fat mass	0.046 (-0.058, 0.149)	-0.041 (-0.196, 0.115)	-0.071 (-0.174, 0.032)	-0.040 (-0.196, 0.116)	-0.102 (-0.202, -0.003)	-0.094 (-0.245, 0.057)	-0.065 (-0.170, 0.039)	-0.076 (-0.234, 0.083)
Total lean mass	0.299 (0.202, 0.397)	0.366 (0.249, 0.433) *	0.096 (-0.005, 0.198)	0.196 (0.076, 0.317)	0.004 (-0.094, 0.103)	0.071 (-0.047, 0.189)	-0.055 (-0.159, 0.048)	-0.046 (-0.170, 0.079)
Metabolic syndrome								
HOMA	-0.121 (-0.220, -0.021)	-0.157 (-0.260, -0.054)	-0.032 (-0.133, 0.069)	-0.013 (-0.118, 0.092)	-0.004 (-0.101, 0.094)	0.021 (-0.081, 0.122)	-0.011 (-0.091, 0.114)	0.026 (-0.081, 0.132)

**Table 3A.** Associations between reproductive and metabolic parameters at 20/21 years of age summarised using standardized beta coefficients and their 95% confidence intervals (CI). All analyses were adjusted for age at 20 years of age, history of cryptorchidism and varicocele ( $\beta_1$ ), coefficients also adjusted for BMI at 20 years of age are shown as ( $\beta_2$ Semen parameters were also adjusted for abstinence period. Unless otherwise specified, data were collected at 20 years of age.

<sup>+</sup>hsCRP>10 are excluded (n=10); effects significant at 0.05 level are shown in bold. \*p<0.001

	S	SCSA	Mor	phology	М	otility
	β1 (95% CI)	β2 (95% CI)	β1 (95% CI)	β <sub>2</sub> (95% CI)	β₁ (95% CI)	β2 (95% CI)
Biochemistry						
Glucose	0.011 (-0.094, 0.116)	0.006 (-0.100, 0.112)	0.042 (-0.064, 0.149)	0.038 (-0.070, 0.146)	0.080 (-0.023, 0.184)	0.081 (-0.024, 0.185)
Triglycerides	-0.065 (-0.170, 0.040)	-0.078 (-0.186, 0.030)	-0.018 (-0.124, 0.089)	-0.027 (-0.137, 0.083)	0.051 (-0.053, 0.154)	0.052 (-0.055, 0.159)
HDL cholesterol	-0.081 (-0.185, 0.024)	-0.077 (-0.186, 0.033)	0.031 (-0.076, 0.138)	0.045 (-0.067, 0.157)	-0.046 (-0.150, 0.057)	-0.048 (-0.157, 0.060)
LDL cholesterol	0.010 (-0.096, 0.115)	0.002 (-0.107, 0.110)	-0.039 (-0.147, 0.068)	-0.049 (-0.159, 0.061)	0.117 (0.014, 0.221)	0.122 (0.016, 0.228)
Iron	-0.011 (-0.117, 0.095)	-0.010 (-0.116, 0.095)	-0.054 (-0.162, 0.053)	-0.054 (-0.161, 0.054)	0.004 (-0.101, 0.108)	0.004 (-0.101, 0.108)
Transferrin	-0.026 (-0.130, 0.079)	-0.030 (-0.136, 0.075)	-0.011 (-0.117, 0.096)	-0.015 (-0.122, 0.093)	-0.055 (-0.159, 0.048)	-0.057(-0.161, 0.047)
Transferrin saturation %	-0.012 (-0.094, 0.118)	0.014 (-0.092, 0.121)	-0.055 (-0.162, 0.052)	-0.053 (-0.161, 0.055)	0.020 (-0.084, 0.125)	0.021 (-0.084, 0.126)
Ferritin	0.047 (-0.059, 0.153)	0.041 (-0.066, 0.149)	-0.065 (-0.173, 0.042)	-0.073 (-0.183, 0.036)	0.065 (-0.040, 0.169)	0.065 (-0.041, 0.172)
Insulin	-0.006 (-0.112, 0.100)	-0.018 (-0.129, 0.092)	0.048 (-0.059, 0.156)	0.042 (-0.070, 0.155)	0.037 (-0.067, 0.141)	0.038 (-0.071, 0.147)
hsCRP <sup>†</sup>	-0.027 (-0.134, 0.081)	-0.040 (-0.152, 0.072)	0.053 (-0.056, 0.162)	0.048 (-0.066, 0.161)	-0.008 (-0.115, 0.098)	-0.011 (-0.122, 0.100)
ALT	0.006 (-0.100, 0.111)	0.005 (-0.115, 0.105)	0.149 (0.043, 0.255)	0.151 (0.040, 0.262)	0.012 (-0.093, 0.116)	0.010 (-0.098, 0.119)
AST	-0.030 (-0.137, 0.077)	-0.045 (-0.158, 0.067)	0.069 (-0.043, 0.181)	0.067 (-0.051, 0.185)	-0.010 (-0.116, 0.096)	-0.014 (-0.125, 0.098)
GGT	-0.044 (-0.149, 0.061)	-0.047 (-0.152, 0.058)	0.122 (0.016, 0.228)	0.120 (0.013, 0.226)	0.024 (-0.079, 0.128)	0.024 (-0.080, 0.128)
Blood pressure						
Systolic	-0.001 (-0.103, 0.101)	-0.016 (-0.125, 0.093)	-0.002 (-0.106, 0.102)	-0.015 (-0.126, 0.096)	0.024 (-0.077, 0.124)	0.024 (-0.084, 0.132)
Diastolic	-0.007 (-0.110, 0.096)	-0.012 (-0.117, 0.092)	-0.049 (-0.154, 0.056)	-0.055 (-0.166, 0.057)	0.059 (-0.043, 0.160)	0.059 (-0.044, 0.161)
Cytokines at 16/17 yrs						
IL18	0.017 (-0.098, 0.132)	0.019 (-0.097, 0.134)	-0.009 (-0.126, 0.108)	-0.007 (-0.124, 0.110)	0.0003 (-0.113, 0.114)	0.001 (-0.113, 0.115)
sTNFR1	-0.030 (-0.145, 0.084)	-0.033 (-0.148, 0.082)	-0.034 (-0.150, 0.083)	-0.036 (-0.153, 0.081)	-0.093 (-0.206, 0.019)	-0.094 (-0.207, 0.019
sTNFR2	0.071 (-0.044, 0.186)	0.070 (-0.046, 0.185)	0.009 (-0.108, 0.127)	0.008 (-0.110, 0.125)	-0.050 (-0.163, 0.064)	-0.050 (-0.164, 0.064
DEXA						
Total fat %	-0.012 (-0.121, 0.097)	-0.064 (-0.209, 0.080)	0.062 (-0.048, 0.172)	0.070 (-0.077, 0.216)	0.052 (-0.056, 0.159)	0.082 (-0.060, 0.225)
Soft tissue fat %	-0.010 (-0.119, 0.099)	-0.061 (-0.205, 0.083)	0.061 (-0.050, 0.171)	0.067 (-0.079, 0.213)	0.051 (-0.057, 0.158)	0.080 (-0.062, 0.222)
Total fat mass	0.007 (-0.102, 0.115)	-0.049 (-0.214, 0.115)	0.045 (-0.065, 0.155)	0.046 (-0.121, 0.213)	0.038 (-0.069, 0.145)	0.074 (-0.088, 0.237)
Total lean mass	0.043 (-0.064, 0.150)	0.033 (-0.096, 0.161)	-0.019 (-0.127, 0.090)	-0.054 (-0.184, 0.077)	-0.038 (-0.144, 0.067)	-0.061 (-0.188, 0.066)
Metabolic syndrome						
HOMA	-0.002 (-0.107, 0.104)	-0.013 (-0.123, 0.097)	0.052 (-0.055, 0.160)	0.047 (-0.065, 0.159)	0.046 (-0.058, 0.151)	0.048 (-0.061, 0.157)

**Table 3B.** (Table 3 continued) Associations between semen parameters and metabolic parameters at 20/21 years of age summarised using standardized beta coefficients and their 95% confidence intervals (CI). All analyses were adjusted for age at 20 years of age, history of cryptorchidism and varicocele ( $\beta_1$ ), coefficients also adjusted for BMI at 20 years of age are shown as ( $\beta_2$ Semen parameters were also adjusted for abstinence period. Unless otherwise specified, data were collected at 20 years of age.

<sup>+</sup>hsCRP>10 are excluded (n=10); effects significant at 0.05 level are shown in bold.

**Table 4.** Associations between serum testicular hormones and gonadotrophins and metabolic parameters at 20/21 years of age summarised using standardized beta coefficients and their 95% confidence intervals (CI). All beta coefficients were adjusted for age at 20 years of age, history of cryptorchidism and varicocele were made in all analyses ( $\beta_1$ ) and separate coefficients are shown with additional adjustment for BMI at 20 years of age ( $\beta_2$ ). Unless otherwise specified, data were collected at 20/21 years of age.

	Testosterone		Inh	В	LH		F	SH
	β1 (95% CI)	β <sub>2</sub> (95% CI)	β <sub>1</sub> (95% CI)	β <sub>2</sub> (95% CI)	β1 (95% CI)	β <sub>2</sub> (95% CI)	β1 (95% CI)	β <sub>2</sub> (95% CI)
Biochemistry				· · · ·	· · ·	· · ·		
Glucose	-0.038 (-0.119, 0.043)	0.002 (-0.077, 0.081)	-0.049 (-0.129, 0.032)	-0.014 (-0.093, 0.065)	-0.018 (-0.098, 0.063)	-0.009 (-0.090, 0.072)	0.003 (-0.077, 0.084)	0.006 (-0.076, 0.087)
Triglycerides	-0.128 (-0.209, -0.048)	-0.068 (-0.148, 0.012)	-0.151 (-0.231, -0.072)*	-0.101 (-0.181, -0.021)	0.036 (-0.045, 0.116)	0.053 (-0.030, 0.135)	0.033 (-0.047, 0.114)	0.039 (-0.044, 0.121)
HDL cholesterol	0.201 (0.122, 0.281) *	0.132 (0.051, 0.213) *	0.095 (0.015, 0.175)	0.027 (-0.054, 0.109)	-0.007 (-0.087, 0.073)	-0.027 (-0.111, 0.057)	-0.024 (-0.104, 0.056)	-0.031 (-0.115, 0.053)
LDL cholesterol	-0.025 (-0.106, 0.057)	0.038 (-0.042, 0.119)	-0.055 (-0.136, 0.026)	-0.002 (-0.083, 0.078)	-0.057 (-0.138, 0.024)	-0.046 (-0.129, 0.037)	-0.020 (-0.101, 0.061)	-0.018 (-0.101, 0.065)
Iron	0.173 (0.092, 0.253) *	0.166 (0.089, 0.243) *	0.014 (-0.067, 0.094)	0.008 (-0.071, 0.086)	-0.045 (-0.126, 0.036)	-0.047 (-0.127, 0.034)	-0.056 (-0.136, 0.025)	-0.056 (0.137, 0.025)
Transferrin	-0.011 (-0.092, 0.070)	0.021 (-0.058, 0.099)	0.014 (-0.066, 0.095)	0.042 (-0.036, 0.121)	-0.031 (-0.111, 0.049)	-0.024 (-0.105, 0.057)	-0.035 (-0.116, 0.045)	-0.034 (-0.115, 0.047)
Transferrin saturation %	0.167 (0.086, 0.247) *	0.149 (0.071, 0.227) *	0.012 (-0.069, 0.093)	-0.004 (-0.083, 0.075)	-0.032 (-0.113, 0.049)	-0.036 (-0.118, 0.045)	-0.039 (-0.120, 0.042)	-0.040 (-0.121, 0.041)
Ferritin	-0.095 (-0.176, -0.013)	-0.048 (-0.128, 0.032)	-0.011 (-0.093, 0.070)	0.031 (-0.049, 0.111)	-0.030 (-0.111, 0.051)	-0.020 (-0.103, 0.062)	-0.025 (-0.106, 0.056)	-0.023 (-0.106, 0.059)
Insulin	-0.241 (-0.320, -0.162)*	-0.177 (-0.258, -0.097)*	-0.211(-0.289, -0.132)*	-0.155(-0.236, -0.074)*	-0.012 (-0.093, 0.069)	0.005 (-0.079, 0.090)	0.072 (-0.009, 0.152)	0.083 (-0.002, 0.167)
hsCRP <sup>†</sup>	-0.249 (-0.329, -0.169)*	-0.187 (-0.268, -0.106)*	-0.024 (-0.106, 0.058)	0.046 (-0.037, 0.129)	-0.073 (-0.155, 0.008)	-0.061 (-0.146, 0.024)	-0.123 (-0.204, -0.042)	-0.129 (-0.214, -0.045)
ALT	-0.116 (-0.197, -0.035)	-0.045 (-0.127, 0.036)	-0.126 (-0.206, -0.045)	-0.067 (-0.148, 0.015)	-0.016 (-0.097, 0.065)	-0.0001 (-0.084, 0.084)	-0.004 (-0.085, 0.077)	-0.001 (-0.085, 0.084)
AST	-0.155 (-0.236, -0.074)	-0.076 (-0.159, 0.008)	-0.149 (-0.230, -0.068)	-0.083 (-0.166, 0.001)	-0.017 (-0.099, 0.065)	0.002 (-0.084, 0.088)	0.038 (-0.044, 0.120)	0.047 (-0.039, 0.133)
GGT	-0.013 (-0.094, 0.068)	0.007 (-0.071, 0.085)	-0.006 (-0.087, 0.074)	0.011 (-0.067, 0.090)	-0.024 (-0.104, 0.057)	-0.028 (-0.135, 0.078)	-0.056 (-0.136, 0.024)	-0.055 (-0.136, 0.025)
Blood pressure								
Systolic	-0.084 (-0.165, -0.004)	0.012 (-0.071, 0.095)	-0.129 (-0.208, -0.049)	-0.053 (-0.136, 0.030)	0.002 (-0.079, 0.082)	0.026 (-0.060, 0.111)	0.037 (-0.043, 0.118)	0.048 (-0.038, 0.134)
Diastolic	-0.086 (-0.167, -0.004)	-0.047 (-0.127, 0.032)	-0.075 (-0.156, 0.006)	-0.042 (-0.122, 0.037)	0.050 (-0.032, 0.131)	0.059 (-0.022, 0.141)	0.062 (-0.019, 0.143)	0.065 (-0.017, 0.147)
Cytokines at 16/17 yrs								
IL18	-0.020 (-0.112, 0.072)	-0.033 (-0.121, 0.056)	0.020 (-0.071, 0.111)	0.009 (-0.079, 0.098)	0.066 (-0.025, 0.157)	0.063 (-0.028, 0.154)	-0.012 (-0.104, 0.079)	-0.013 (-0.104, 0.978)
sTNFR1	-0.027 (-0.119, 0.064)	-0.010 (-0.099, 0.078)	-0.128 (-0.218, -0.038)	-0.114 (-0.202, -0.026)	0.108 (0.018, 0.198)	0.112 (0.022, 0.202)	0.153 (0.063, 0.242)*	0.154 (0.064, 0.244)*
sTNFR2	-0.004 (-0.095, 0.088)	0.011 (-0.078, 0.099)	-0.011 (-0.102, 0.079)	-0.001 (-0.087, 0.090)	0.039 (-0.051, 0.130)	0.043 (-0.048, 0.134)	0.037 (-0.054, 0.128)	0.038 (-0.053, 0.129)
DEXA								
Total fat %	-0.243 (-0.327, -0.159)*	-0.105 (-0.214, 0.005)	-0.137 (-0.222, -0.052)	0.036 (-0.074, 0.146)	-0.120 (-0.205, -0.035)	-0.139 (-0.252, -0.027)	-0.076 (-0.161, 0.010)	-0.115 (-0.228, -0.002)
Soft tissue fat %	-0.241(-0.325, -0.157)*	-0.103 (-0.212, 0.006)	-0.139 (-0.224, -0.054)	0.031 (-0.079, 0.140)	-0.121 (-0.206, -0.036)	-0.140 (-0.252, -0.028)	-0.074 (-0.159, 0.012)	-0.111 (-0.223, 0.001)
Total fat mass	-0.271 (-0.354, -0.188)*	-0.143 (-0.268, -0.019)	-0.175 (-0.259, -0.091)*	0.010 (-0.114, 0.135)	-0.112 (-0.197, -0.027)	-0.150 (-0.277, -0.022)	-0.058 (-0.143, 0.028)	-0.106 (-0.234, 0.022)
Total lean mass	-0.038 (-0.122, 0.047)	0.167 (0.070, 0.265)*	-0.139 (-0.222, -0.055)*	-0.009 (-0.108, 0.089)	0.052 (-0.032, 0.136)	0.124 (0.023, 0.224)	0.086 (0.002, 0.170)	0.136 (0.036, 0.237)
Metabolic syndrome								
HOMA	-0.237 (-0.341, -0.133)*	-0.172 (-0.278, -0.066)	-0.203 (-0.282, -0.124)*	-0.149(-0.230, -0.068)*	-0.016 (-0.097, 0.065)	0.001 (-0.083, 0.085)	0.069 (-0.011, 0.150)	0.080 (-0.004, 0.164)

<sup>†</sup>hsCRP>10 are excluded (n=10); effects significant at 0.05 level are shown in bold. \*p = or <0.001

**Table 5.** Comparison of testicular volume, semen parameters and serum hormones by metabolic clusters, all assessments made at 20 years of age in the top part of the table, and metabolic cluster analysis and associations at 17 years of age are listed in the lower part of the table. Data is represented as Median (IQR, R) and Mean (SD) as appropriate.

Cluster parameters at 20 years of age	N Hig	High risk at 20 years of age [Mean (SD)]	NLow	Low risk at 20 years of age [Mean (SD)]	p-value
	h				
Systolic blood pressure (mm/Hg)	43	130.8 (10.6)	342	121.8 (12.2)	<0.001
İnsulin (µu/ml)	43	14.2 (12.8)	342	3.2 (2.2)	<0.001
Triglycerides (mmol/l)	43	1.8 (1.1)	342	1.0 (0.4)	<0.001
Waist circumference (cm)	43	100.2 (12.9)	342	80.2 (7.6)	<0.001

# Metabolic cluster at 20 years of age

Testicular function		High risk at 20 years of age		Low risk at 20 years of age	
assessment at 20 years		[Median (IQR, R)]		[Median (IQR, Ř)]	
Testicular volume (ml)	42	14.7 (12.3-16.9, 9.0-23.8)	325	15.2 (13.0-17.4, 7.6-28.4)	0.574
Semen parameters					
Volume (ml)	34	2.7 (1.9-4.0, 0.9-7.2)	303	2.8 (1.9-3.7, 0.1-11.0)	0.979
Total sperm output (M)	34	115.3 (51.0-194.0, 0.0-551.8)	303	113.4 (50.6-207.0, 0.0-927.5)	0.738
Sperm concentration	34	42.5 (19.4-70.5, 0-142)	303	46 (23-73, 0-220)	0.663
(M/mL)					
SCSA (%)	32	2.5 (1.5-4.7, 0.6-10.8)	298	3.1 (1.9-5.2, 0.2-30.0)	0.106
Morphology (N, %)	32	5.5 (3.6-9.0, 3-17)	294	5 (3-7, 0-18)	0.144
Motility (a + b, %)	33	58.0 (43.5-70.5, 19-86)	300	58 (44-67, 1-88)	0.773
Serum hormones					
Testosterone (ng/ml)	75	3.6 (3.0-4.0, 1.1-6.5)	522	4.8 (3.8-5.9, 1.3-10.3)	<0.001
LH (IU/I)	76	6.7 (7.6-12.8, 5.2-19.3)	522	10.5 (8.3-13.1, 2.3-28.4)	0.097
FSH (IU/I)	76	4.4 (2.9-6.8, 0.8-25.8)	522	4.3 (3.0-6.1, 0.6-39.5)	0.492
InhB (pg/ml)	76	167.9 (132.1-217.0, 28.9-389.3)	523	223.7 (180.6-272.9, 4.5-543.9)	<0.001

# Metabolic cluster at 17 years of age

Testicular function		High risk at 16/17 yrs of age		Low risk at 16/17 yrs of age	
assessment at 20 years		[Median (IQR, R)]		[Median (IQR, R)]	
Testicular volume (ml)	39	15.6 (13.3-17.5, 10.1-23.2)	249	14.7 (12.6-17.1, 8.0-28.4)	0.215
Semen parameters					
Volume (ml)	37	2.5 (1.6-3.6, 0.3-11.0)	227	2.8 (2.0-3.6, 0.7-7.5)	0.347
Total sperm output (M)	37	110.7 (52.2-288.9, 0.0-592.2)	227	122.2 (56.0-217.6, 0.0-927.5)	0.711
Sperm concentration	37	50 (26.5-88.5, 0-220)	227	47 (23-71, 0-210)	0.280
(M/mL)					
SCSA (%)	35	3.6 (1.8-6.5, 0.7-30)	222	3.3 (1.9-5.5, 0.2-19.0)	0.416
Morphology (N, %)	35	5.0 (3.0-7.0, 0.5-17)	219	4.5 (3.0-7.0, 0.5-18.0)	0.782
Motility (a + b, %)	36	51.0 (38.5-65.8, 7.0-88.0)	224	59.0 (43.3-68.0, 7.0-88.0)	0.170
Serum hormones					
Testosterone (ng/ml)	67	4.0 (3.2-4.9, 1.6-7.2)	356	4.9 (3.6-6.0, 1.8-9.9)	<0.001
LH (IU/I)	68	10.1 (7.8-13.9, 5.4-19.8)	357	10.6 (8.6-13.2, 4.3-28.4)	0.425
FSH (IU/I)	68	4.4 (3.4-6.8, 1.1-14.3)	357	4.3 (3.0-6.2, 0.8-39.5)	0.285
InhB (pg/ml)	68	193.2 (144.8-226.5, 48.7-389.3)	357	221.9 (180.3-269.0, 56.7-	<0.001
				543.9)	

Testicular function	N <sub>IR</sub>	IR (HOMA>4)	Normal	Normal (HOMA≤4)	p-value
Testicular volume (ml)	14	12 8 (11 1-14 7 10 0-16 9)	359	15 2 (13 0-17 4 7 6-28 4)	0.010
Sperm parameters	14	12.0 (11.1-14.7, 10.0-10.0)	000	10.2 (10.0-11.4, 1.0-20.4)	0.010
Semen volume (ml)	13	2.6 (1.5-3.6. 0.9-4.2)	326	2.8 (1.9-3.8. 0.1-11.0)	0.320
Total sperm output (M)	13	136.8 (81.0-253.9, 0.0-	326	110.6 (50.6-206.7, 0.0-	0.459
		383.4)		927.5)	
Sperm concentration (M/ml)	13	64.0 (30.0-88.5, 0-160)	326	44.5 (22.0-70.3, 0-220)	0.293
SCSA (%)	12	2.8 (1.5-5.4, 1.4-10.8)	320	3.1 (1.8-5.2, 0.2-30.0)	0.654
Sperm morphology (N, %)	12	5.5 (3.3-10.0, 3-17)	316	5 (3-7, 0-18)	0.402
Sperm motility (a + b, %)	12	63 (47.3-75.8, 26-79)	323	58 (43-67, 1-88)	0.330
Serum hormone concentrations					
Testosterone (ng/ml)	24	3.2 (2.6-4.0, 1.1-5.3)	583	4.6 (3.7-5.9, 1.3-10.3)	<0.001
LH (IU/I)	24	10.6 (8.2-13.3, 6.3-17.1)	584	10.5 (8.3-13.0, 2.3-28.4)	0.903
FSH (IU/I)	24	6.1 (3.4-7.9, 1.1-14.3)	584	4.3 (3.0-6.1, 0.6-39.5)	0.046
InhB (pg/ml)	24	172.4 (130.0-213.4, 54.8-	585	217.8 (174.0-267.6, 4.5-	0.001
		389.3)		543.9)	

**Table 6.** Comparison of testicular volume, semen parameters and serum testicular hormones and gonadotrophins by HOMA-IR, all assessments made at 20 years of age. Data is represented as Median (IQR, R).

HOMA = homeostasis model assessment insulin (Fasting insulin  $[\mu u/ml] \times Fasting glucose [mM]/22.5$ ).