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BRIEF REPORT



Tropomyosin autoantibodies associated with checkpoint inhibitor myositis

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ABSTRACT

This brief report details the measurement and identification of IgA antibodies to tropomyosin in a case of presumed ocular myositis with paraspinal myositis in a patient with metastatic uveal melanoma treated with checkpoint inhibitors. High-throughput functional protein microarray analysis and pathway analysis was conducted to identify IgG and IgA antibodies of interest. Antibody levels were compared to generic antibody screening results and levels of the antibodies in a cohort of melanoma patients without myositis (n = 100) at baseline prior to undergoing immunotherapy. The finding of specific muscle antibodies in this clinical case indicates the pathogenic potential of anti-tropomyosin IgA in the development of checkpoint inhibitor associated myositis and requires further investigation.

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KEYWORDS

Tropomyosin; myositis; ocular melanoma; autoantibody; immunotherapy

Brief report

We recently reported a case of presumed ocular myositis with paraspinal myositis in a patient with metastatic uveal melanoma treated with checkpoint inhibitors Pembrolizumab and Ipilimumab/Nivolumab.¹ Conventional IgG autoantibody screening in this case was negative (anti-AChR, anti-ganglioside, myositis specific antibodies and anti-MuSK). The patient's level of creatine kinase (2452 U/l) and troponin I (0.36 micrograms/l) was elevated and an MRI of the cervical muscle was consistent with myositis.

Serum was taken, two days after hospital admission and prior to steroid and intravenous immunoglobulin infusion that achieved complete clinical response. Unfortunately, the patient passed away at 12 months post immunotherapy commencement due to cerebral metastases and a further trial of radiotherapy and other checkpoint inhibitors. Retrospective informed consent was obtained from the legal representative of the deceased and is available upon request. The serum was analyzed by microarray for IgG and IgA binding efficiencies (Supplementary Tables 1 and 2, respectively) to greater than 21000 antigens (>81% of the human proteome) using the HuProtTM microarray (CDI Laboratories, USA). The sample was probed on the array at a 1:1000 dilution and analyzed for Z scores, which identified 84 and 172 positive hits (Z score > 3) for IgG and IgA respectively. Log₂ fold changes (log₂FC) were calculated by dividing the case signal intensity for each antigen by the mean case signal intensity of all antigens or the median signal intensity of all antigens across the cohort of other metastatic melanoma samples. In particular, the microarray results demonstrated high levels of IgA antibodies to tropomyosin (TPM) isoforms 1, 2 and 3 while IgG antibodies to these antigens were found to be negative. Tropomyosin 3 is essential for melanoma metastasis, enabling

pseudopodium and invadopodium formation.² Sera from a group of metastatic melanoma patients without myositis (n = 100) at baseline prior to undergoing immunotherapy who were also screened on the array (unpublished, Edith Cowan University Ethics Committee application number 18957) were negative to these antigens (Figure 1).

The log₂ IgA and IgG data for all antigens in this case study was then uploaded to Advaita Bio's iPathwayGuide (<http://www.advaitabio.com/ipathwayguide>) software to determine major pathways enriched by the differentially elevated antibodies (*p*-value < 0.05 and a |log₂FC| > 3.47 (Z score > 3)). A detailed description of the pathway analysis approach utilized is described elsewhere.³ Using this software, a total of 97 elevated antibodies were identified and 19 biological pathways were found to be significantly impacted, with cardiac muscle contraction, carbon metabolism and hypertrophic cardiomyopathy pathways showing the most significant involvement in the case study's IgA binding profile (*p* = 5.188 × 10⁻⁴, *p* = .004, *p* = .007, respectively). Similar to the microarray results, the most significant identified biological components associated with the case IgA binding profile, included muscle thin filament tropomyosin, striated muscle thin filament and myofilament (*p* = 7.4 × 10⁻⁷, *p* = 8.1 × 10⁻⁵, *p* = 9.9 × 10⁻⁵, respectively).

Myositis is an autoimmune/antibody-mediated condition and several myositis-related and -associated autoantibodies have been identified to date.^{4,5} The regionalization of the myositis (paraspinal, ocular and myocardial) in this case is interesting as in a mouse model of muscular dystrophy, deletion of a tropomyosin 3 isoform (Tpm3.1) caused muscle disease in a similar distribution.⁶ Garaud et al.⁷ performed microarray antibody analysis in breast cancer, with sera and

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 Supplemental data for this article can be accessed [here](#).

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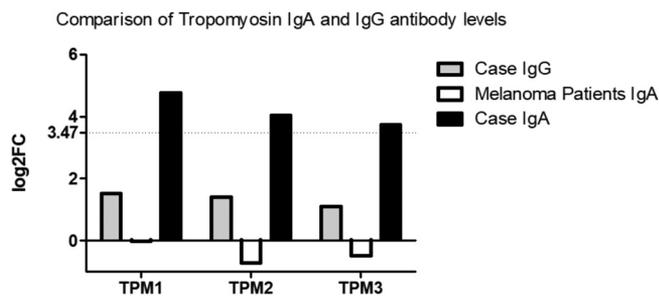


Figure 1. Comparison of IgA and IgG levels to tropomyosin isoforms 1, 2 and 3 in the reported clinical case study. IgA levels from the sera of a group of metastatic melanoma patients without myositis ($n = 100$) at baseline prior to undergoing immunotherapy are also illustrated. Levels are presented as Log2 fold changes (\log_2FC) and were calculated by dividing the case signal intensity for each antigen by the mean case signal intensity of all antigens or the median signal intensity of all antigens across the cohort of other metastatic melanoma samples. The horizontal line at $\log_2FC = 3.47$ represents the cutoff where the case Z score > 3 , indicating a positive signal intensity.

breast tissue displaying high levels of tumor specific IgA to tumor antigens, including cancer/testis antigen 1B (CTAG1B) and ankyrin repeat domain 30B like protein (ANKRD30BL). These were not related to tumor progression or survival. There was however, a correlation with the development of tertiary lymphoid structures within the tumor suggesting local IgA production. B cell infiltration in tumors is rare but B cell activation does occur in primary, secondary and tertiary lymphoid structures and antibodies may play a role in tumor destruction or progression. Whilst IgA is unable to directly activate the complement pathway, it can do so via the mannose lectin pathway. It is now recognized that monomeric IgA opsonised on cell membranes is able to cause apoptosis and necrosis by binding to the Fc α R1 receptor (CD89) on neutrophils.⁸ The exact mechanism of subsequent tissue damage is under debate and a new novel process called trogoptosis has been suggested.⁹

In summary, the finding of specific muscle antibodies in this case may have played a role in the checkpoint induced myositis and further studies are required to elucidate the pathogenic potential of anti-tropomyosin IgA antibodies.

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Author contributions

| Name | Location | Role | Contribution |
|----------------------|-------------------------------|--------|--|
| Pauline Zaenker, PhD | Edith Cowan University, Perth | Author | Design and conceptualized study; Data collection and analysis, drafted the manuscript for intellectual content |

(Continued)

| Name | Location | Role | Contribution |
|----------------------|---|--------|--|
| Pauline Zaenker, PhD | Edith Cowan University, Perth | Author | Design and conceptualized study; Data collection and analysis, drafted the manuscript for intellectual content |
| David Prentice, MD | St John of God Midland Public Hospital, Midland | Author | Design and conceptualized study; Data collection and analysis, Drafting and revision for intellectual content |
| Melanie Ziman, PhD | Edith Cowan University and the University of Western Australia, Perth | Author | Design and conceptualized study; Drafting and revision for intellectual content |

Disclosure statement

The authors of the manuscript declare no conflict of interest.

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